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Seaweeds as nutraceuticals for health and nutrition

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ABSTRACT

Throughout human history, seaweeds have been used as food, folk remedies, dyes, and as mineral-rich fertilisers. Seaweeds as nutraceuticals or functional foods with dietary benefits beyond their fundamental macronutrient content are now a major research and industrial development concept. The occurrence of dietary and lifestyle-related diseases, notably type 2 diabetes, obesity, cancer, and metabolic syndrome has become a health epidemic in developed countries. Global epidemiological studies have shown that countries where seaweed is consumed on a regular basis have significantly fewer instances of obesity and dietary-related disease. This review outlines recent developments in seaweed applications for human health from an epidemiological perspective and as a functional food ingredient.

KEYWORDS

Functional foods; Human health; Lifestyle-related disease; Nutraceuticals; Seaweed

Abbreviations:

B.N.F. British Nutrition Foundation; C.D.C. Centers for Disease Control and Prevention U.S.A.; C.M.M.M. Chinese Marine Materia Medica; DPPH 2,2-diphenyl-1-picrylhydrazyl; GAE gallic acid equivalents; LDL low-density lipoprotein; ORAC oxygen radical absorbance capacity; PUFA polyunsaturated fatty acid; RNI adult reference nutrient intake; W.H.O. World Health Organization.

INTRODUCTION

Seaweeds are marine, photosynthetic algae which are abundant in every ocean. There are three main classes or phyla of seaweed: Phaeophyceae (brown algae), Rhodophyta (red algae), and Chlorophyta (green algae). Thousands of species comprise each phylum (Rindi *et al.* 2012; Guiry & Guiry 2019). Seaweeds have been used traditionally as food, folk remedies, dyes, and fertilisers. With the development of mass food manufacturing in the early 1900's, seaweeds components were harnessed industrially. Hydrocolloids, such as alginate, carrageenan and agar, are still the most commonly used components for their gelling properties in foods, pharmaceutical and biotechnological applications (Rhein-Knudsen *et al.* 2015; Zollman 2019). The suitability of seaweed for bio-diesel production has been explored for developing green fuel alternatives (Chye *et al.* 2018; Abomohra *et al.* 2018).

In the last three decades interest has grown in seaweeds as nutraceuticals or functional foods, with dietary benefits beyond their macronutrient content. In addition, seaweed has been mined for metabolites with biological activity, to produce therapeutic products (Davis & Vasanthi 2011; Zerrifi *et al.* 2018). The paradox of obesity concurrent with mineral and vitamin deficiency has become a health epidemic in developed regions such as Europe, the U.S.A., and Australia (W.H.O. 2019). At the same time, the occurrence of dietary and lifestyle-related diseases, notably type 2 diabetes, cancer, and metabolic syndrome has increased. Global dietary studies have found that countries where seaweed is consumed on a regular basis have significantly less obesity and dietary-related diseases (Iso 2011; Nanri *et al.* 2017). Although a number of dietary factors may be involved in this inverse relationship, the study of seaweed alone for its beneficial dietary and medicinal effects has warranted investigation. This review outlines recent developments in

seaweed applications for human health from an epidemiological perspective and as a functional food ingredient.

Functional food applications of seaweed

MICRO- AND MACRONUTRIENT PROFILE: Seaweed has been foraged and consumed by coastal communities worldwide since the beginning of human civilisation (Dillehay *et al.* 2008). Incorporating seaweeds or their extracts into foods to improve nutritional properties is a recent practice, prompted by improved understanding of dietary sciences and the nutrient-dense nature of algae.

MINERALS AND VITAMINS: Some seaweeds contain 10 to 100 times more minerals and vitamins per dry mass than terrestrial plants or animal-derived foods (Rupérez 2002). These include fat- and water-soluble vitamins A, D, E, K, C, B₁, B₂, B₉, B₁₂ and essential minerals, calcium, iron, iodine, magnesium, phosphorus, potassium, zinc, copper, manganese, selenium, and fluoride (Misurcova 2011; Qin 2018). Content varies among species. For example, a study of five brown, eight red, and eight green seaweeds from northern European waters found that total tocopherol (vitamin E) content ranged from 1.6-122 mg kg⁻¹ in brown, 10-26 mg kg⁻¹ in red, and 8.8-12.0 mg kg⁻¹ in green species (dry mass) (Biancarosa *et al.* 2018). From a portion perspective, taking 8 g (dry mass) as a typical serving size, many seaweeds perform better than plant and animal foods in terms of adult reference nutrient intake (RNI) (Astorga-España *et al.* 2015). For example, the British Nutrition Foundation recommends 8.7 mg iron/day for adult males (B.N.F. 2016). The red seaweed, *Palmaria palmata* (see Table 1 for authorities, common names and higher taxonomic position), contains on average 6.4 mg of iron per 8 g serving, compared to only 1.2-3.1 mg in a 100 g portion of lean beef (Branscheid & Judas 2011). Similarly, 8 g of the green seaweed, *Ulva*

lactuca, contains on average 260 mg of calcium or 37% of the RNI, while 8 g of cheddar cheese provides on average 5% of the RNI (MacArtain *et al.* 2007; Finglas *et al.* 2015).

Table 1. List of algal species mentioned in the text, by taxonomic group with common names. Note: The initial name is that used in the cited literature; the name after the equal sign is the one currently accepted. Synonymy and common names (where available) follow AlgaeBase (Guiry & Guiry 2019).

Phylum or Class	Species	Selected common names
Cyanobacteria	<i>Spirulina platensis</i> (Gomont) Geitler = <i>Arthrospira platensis</i> Gomont	Spirulina
Chlorophyta (green algae)	<i>Chaetomorpha linum</i> (O.F.Müller) Kützinger	Warakuzumo, flax brick weed (U.K.)
	<i>Chlorella vulgaris</i> Beyerinck	
	<i>Ulva lactuca</i> Linnaeus	Sea lettuce
	<i>Enteromorpha prolifera</i> O.F.Müller = <i>Ulva prolifera</i> (O.F.Müller) J.Agardh	
Rhodophyta (red algae)	<i>Euclima cottonii</i> Weber van Bosse = <i>Kappaphycus alvarezii</i> (Doty) Doty ex Silva	Elkhorn sea moss
	<i>Gracilariopsis lemaneiformis</i> (Bory de Saint-Vincent) E.Y.Dawson, Acleto & Foldvik	
	<i>Palmaria palmata</i> (Linnaeus) F.Weber & D.Mohr	Dulse
	<i>Porphyra/Pyropia</i>	Nori, gim, laver
	<i>Porphyra umbilicalis</i> Kützinger	Nori, laver
	<i>Pyropia columbina</i> (Montagne) W.A.Nelson	Nori, southern laver
Phaeophyceae (brown algae)	<i>Ascophyllum nodosum</i> (Linnaeus) Le Jolis	Knobbed wrack
	<i>Cystoseira barbata</i> (Stackhouse) C.Agardh	
	<i>Durvillaea antarctica</i> (Chamisso) Hariot	Cochayugo
	<i>Ecklonia cava</i> Kjellman	Kajime
	<i>Fucus spiralis</i> Linnaeus	Spiral wrack
	<i>Fucus vesiculosus</i> Linnaeus	Bladder wrack
	<i>Himanthalia elongata</i> (Linnaeus) S.F.Gray	Sea spaghetti
	<i>Laminaria hyperborea</i> (Gunnerus) Foslie	Kelpie
	<i>Laminaria japonica</i> Areschoug = <i>Saccharina japonica</i> (Areschoug) C.E.Lane, C.Mayes, Druehl & G.W.Saunders	Sea Tangle, dasima
	<i>Macrocystis pyrifera</i> (Linnaeus) C.Agardh	Giant kelp
	<i>Saccharina latissima</i> (Linnaeus) C.E.Lane, C.Mayes, Druehl & G.W.Saunders	Sugar kelp, kombu
	<i>Saccorhiza polyschides</i> (Lightfoot) Batters	Furbelows
	<i>Sargassum thunbergii</i> (Mertens ex Roth) Kuntze	Sargassum
	<i>Sargassum fusiforme</i> (Harvey) Setchell	Hijiki
	<i>Sargassum vulgare</i> C.Agardh nom. illeg.	Beerentang
<i>Sargassum crassifolium</i> J.Agardh = <i>Sargassum aquifolium</i> (Turner) C.Agardh	Binder's Sargassum weed	
<i>Undaria pinnatifida</i> (Harvey) Suringar	Wakame	

PROTEIN: Protein constitutes 5-47% of seaweed dry mass. Red seaweeds have the greatest protein content, while green have less, and brown the least (Černá 2011). Of the total amino acids (aa) in seaweeds, approximately 42-48% are aa (Wong & Cheung 2000). In terms of aa score (on a scale of 0.0-1.0), where egg protein has a score of 1.0, most seaweeds have a higher score than all plant-based proteins, with the exception of soy, which has a score of 1.0. For example *Undaria pinnatifida* has an amino acid score of 1.0, equal to that of egg and soy, *Pyropia/Porphyra* 0.91, and *Laminaria saccharina* 0.82 (Murata & Nakazoe (2001). However, the high polyphenolic content of seaweeds can reduce the digestibility of algal proteins, giving a slightly lower score on

the protein digestibility-corrected aa scale (Wong & Cheung 2001). Despite this, seaweeds still represent a viable alternative to animal-derived protein, if other high-aa scoring vegan foods, such as soy mycoprotein, are included in the diet.

POLYSACCHARIDES: Total polysaccharide or sugar content of seaweeds, ranges from 4% to 76% (dry mass) (Paniagua-Michel *et al.* 2014). Cellulose is an indigestible, non-nutritive polysaccharide that gives structure to cell walls of many seaweeds and constitutes from 2-10% of total polysaccharides. Digestible polysaccharides differ between phyla. Alginates, fucoidans, and the laminarin are the main polysaccharides in brown algae (Rodrigues *et al.* 2015); carrageenans and agarans occur in red algae; and ulvans in green algae (Jiao *et al.* 2011). Most algal polysaccharides are non-starchy fibre, which help balance normal blood-glucose levels, and can contribute to the RNI of 30 g fibre day⁻¹ (B.N.F. 2016). The structural chemistry of algal fibres differs from those found in terrestrial plants. This gives them functional and bioactive properties not found in land-based fibre sources (Jimenez-Escrig & Goñi 1999).

LIPIDS: Total lipid content of seaweeds ranges from 0.60–4.14% (Rodrigues *et al.* 2015; El Maghraby & Fakhry 2015). Most algal lipids are polyunsaturated, composed of (*n*-3, or omega-3) fatty acids such as docosahexaenoic and eicosapentaenoic acid. Linoleic and arachidonic acids are the most common algal mono-unsaturated (*n*-6, or omega-6) fatty acids (Belattmania *et al.* 2018). Principle saturated fatty acids include palmitic and myristic acids. From a dietary perspective, both *n*-6 and *n*-3 fatty acids are essential, however, consuming them in an imbalanced ratio can result in chronic inflammatory diseases such as obesity, rheumatoid arthritis, non-alcoholic fatty liver, and cardiovascular disease (Patterson *et al.* 2012). The ratio of *n*-6:*n*-3 consumption in developed,

Western countries has risen to approximately 20:1 in the last two decades (Logan 2003; Husted & Bouzinova 2016). A ratio between 2.5:1 and 4:1 ($n-6:n-3$) is generally recommended to prevent chronic diseases associated with excess $n-6$ monounsaturated fat consumption (Simopoulos 2016). The $n-6:n-3$ ratio of fatty acids in seaweeds is within this low ratio (Biancarosa *et al.* 2018), making them excellent dietary lipid sources (Dawczynski *et al.* 2007).

Other nutrient compounds unique to seaweeds, such as fucoxanthin, phycobiliproteins, phlorotannins, and sulphated polysaccharides have therapeutic effects beyond basic nutrition (Mysliwa-Kurziel & Solymosi 2017; Pérez *et al.* 2016). These will be discussed below.

INCORPORATION IN FOOD PRODUCTS

The nutrient-dense nature of seaweeds makes them excellent candidates for functional food use. In addition, their physical ability to emulsify and retain water enhances their techno-functional properties as food ingredients. Examples of nutritional, structural, antimicrobial, organoleptic, and shelf-life enhancement of meat or plant-based food products by the addition of seaweed are outlined below.

MEAT-BASED PRODUCTS: Meat is part of the staple diet of billions of people worldwide and is a good source of protein, bioavailable iron, zinc, selenium, magnesium, and vitamins B, D, and A. However, meats, particularly pork and beef, contain substantial levels of saturated fat which can raise LDL cholesterol levels. Meat-derived products such as burgers, frankfurters, salami, and deli-sliced meats also contain salt as a necessary ingredient for preservation, flavour and texture enhancement. Some manufacturers also add artificial flavour enhancers and preservatives such as monosodium glutamate and sodium nitrate which have been linked to cancer and other disorders

(Ramesh & Muthuraman 2018). Regular consumption of saturated fat, salt, and artificial additives has contributed to disorders such as heart disease and obesity worldwide. The search for compounds to reduce salt, saturated fat, and eliminate artificial additives in meat products has led to use of seaweed components as healthy functional ingredients. The incorporation of a variety of seaweeds has produced several meat products of high consumer acceptance with significantly lower levels of saturated fat and salt, no artificial additives, plus increased fibre and polyunsaturated fat content (Cofrades *et al.* 2017). In addition, polyphenolic compounds, selenium, and vitamins A, C, and E in seaweed act as natural antioxidants and preservatives within the meat matrix.

Using *Undaria pinnatifida*, *Porphyra umbilicalis*, and *Himanthalia elongata*, López-López *et al.* (2009) incorporated 5.6% dried, milled seaweed into pork frankfurters, beef burgers, and restructured poultry steaks with consumer sensory acceptance. The seaweeds significantly decreased the thrombogenic index of the products' fatty acid profile, by increasing the ratio of *n*-3 to *n*-6 polyunsaturated fats. Sodium was significantly decreased, while calcium, magnesium and manganese, vitamin K, and soluble polyphenolic compounds levels were also increased. Total polyphenolic content was highest in meats enriched with *H. elongata* (25.7 mg gallic acid equivalents (GAE)/g), followed by *P. umbilicalis* (21.7 mg GAE g⁻¹) and *U. pinnatifida* (0.82 mg GAE g⁻¹). The same trend was seen in antioxidant capacity – measured by the ferric reducing antioxidant power assay – where meat enriched with *H. elongata* had an antioxidant capacity of 3.69 µmol Trolox equivalents (TE) g⁻¹, followed by *P. umbilicalis* (1.18 µmol TE g⁻¹) and *U. pinnatifida* (1.09 µmol TE g⁻¹). The seaweed-enriched meats were later tested in animal models and significantly improved lipoprotein metabolism was found, suggesting that these products

could be developed as a hyper-cholesterolaemic in human dietary interventions (Olivero-David *et al.* 2011).

Himanthalia elongata was also used to enrich beef patties (Cox & Abu-Ghannam 2013); however, the seaweed was not used in dried powder as per López-López *et al.* (2009). Instead, whole, dried *H. elongata* was blanched in water for 20 min at 80 °C before being blended to a paste, and combined with fresh minced beef (10–40% w/w). Unlike most studies which report the highest sensory acceptability in products with the lowest concentrations of seaweed, the patties containing 40% seaweed were rated by the sensory panel as having the best overall acceptability in terms of aroma, texture, taste and appearance. Throughout chilled storage, all chemical, physical, sensory and microbial parameters were significantly improved by addition of *H. elongata*. Fibre increased from zero in the control to 1.64 g total dietary fibre per 100 g (fresh weight). Water-retention is a useful property of algal fibre where fat content is reduced, as it retains moisture in foods. The beef used in this study had a fat content of 14%. Replacing 40% of the meat with seaweed reduced the fat content to just 8.4%, while maintaining full sensory acceptance by the panel of judges. In addition, moisture loss (by mass) during cooking was reduced by 6.4%. After 30 days at 4 °C, the total viable microbiological plate count in the cooked, control burger was 5.41 log CFU g⁻¹ compared to zero CFU g⁻¹ in the 20%, 30% and 40% seaweed-enriched burgers. Growth occurred (1.09 log CFU g⁻¹) only in the 10% seaweed burgers after 30 days. Final lipid oxidation levels were 45% lower in the 40% seaweed-enriched burgers (0.61 mg malondialdehyde kg⁻¹) compared to 1.12 mg MDA kg⁻¹ in the control. Total phenolic content and DPPH radical scavenging activity were zero in the control sample, and as high as 28.11 mg GAE 100 g⁻¹ and 52.32% respectively (fresh weight) in the 40% seaweed-enriched burgers. Tenderness also improved significantly as shown by via Instron measurement of firmness/hardness after

storage for 30 days. Hardness was measured as 40.23 N mm⁻¹ in the control sample, compared to only 21.33 N mm⁻¹ in the 40% seaweed-enriched burgers, indicating that they were almost 47% more tender.

Sellimi *et al.* (2017) improved the functional and nutritional attributes of cured turkey meat sausages with the addition of fucoxanthin, extracted from *Cystoseira barbata*. Over 15 days, there was a significant increase in antioxidant capacity, colour stability, inhibition of lipid peroxidation, and angiotensin-I converting enzyme which raises blood pressure. There was also a 47% reduction in the amount of sodium nitrite required as an anti-bacterial, from 150 ppm to 80 ppm. Only 0.04% (total ingredient composition) of fucoxanthin was required to achieve these improvements, while still maintaining consumer sensory acceptance.

These nutritional and technological quality enhancements, combined with sensory acceptance, were achieved with a cost effective, sustainable ingredient and minimal preparation, making the addition of seaweed a viable option for meat product enrichment. Seaweed as a functional food ingredient can allow consumers to continue eating meat products while reducing their risk of adverse health effects.

PLANT-BASED PRODUCTS: Aside from unprocessed vegetables, fruit, pulses, and rice, the majority of plant-based (processed) products consumed globally are derived from grains, and constitute part of the daily diet of almost all human populations (Stephen *et al.* 2017). These grain- or cereal-based products are consumed as thousands of different varieties of bread, pasta, noodles, and pastries. Since the majority of these foods are produced using refined white flours, the fibre, protein, mineral and vitamin content is poor, while starch content is high. Therefore, enrichment of cereal-based products with high fibre, nutrient-dense functional ingredients such as seaweed

has the potential to increase the dietary intake of essential nutrients. Several studies have successfully incorporated seaweed and their extracts into cereal-based products.

Kumoro *et al.* (2016) improved the fibre, protein, lipid, and mineral profile of wheat flour noodles with the addition of dried, milled *Eucheuma cottonii*. In addition to seaweed, two dried microalgal powders of *Spirulina platensis* and *Chlorella vulgaris* were also added. The control noodles were made using 100 g of white Indonesian wheat flour, eggs, water, and salt. Dried *E. cottonii* was added alone, or combined with *S. platensis* and *C. vulgaris*, in ratios from 10-30 g, with wheat flour of 90-70 g as required to produce a total dry mass of 100 g for each batch of noodles. In terms of nutritional improvement and sensory acceptance, the optimum composition was 90 g wheat flour, with 5 g *E. cottonii* and 5 g *S. platensis*. This produced noodles with the most protein, lowest fat content, and greatest consumer acceptability. Total carbohydrate content (mostly starch), was reduced from 78% to 68% in control noodles; and crude fibre increased by 125%, from 1.72 g per 100 g in the control to 3.87 g per 100 g in algal enriched samples. This was due to the high fibre content (25%) of dried *E. cottonii* compared to only 0.5% fibre in wheat flour. Protein increased by 87% (from 9.34 g per 100 g to 17.53 g per 100 g), while total mineral (ash) content almost tripled, from 1.01% to 3.01%. Organoleptic parameters of texture, flavour, aroma, and colour scored positively, similar to the wheat flour-only control. Some parameters were in fact higher in the 10% *E. cottonii* and *S. platensis* enriched samples, with panellists showing a preference for the light green colour and savoury aroma of the noodles.

Arufe *et al.* (2018) harnessed the techno-functional properties of *Fucus vesiculosus* to improve the antioxidant profile of white bread while maintaining acceptable density and crumb texture. *Fucus vesiculosus* was dried and milled to a powder. A control recipe of white wheat flour (300 g), fresh yeast (6 g), tap water (192 g), salt (5.4 g) and ascorbic acid (0.006 g) was used. The

algal powder was added as 2-8% (6-24 g) of the total flour mass. After all other ingredients were added and kneaded together the dough was proofed (27 °C, 35 min). Dough was divided and proofed again (27 °C, 70 min), then baked (245 °C, 21 min). The proofing behaviour and rheological properties of the doughs were measured to determine the maximum percentage of *F. vesiculosus* that could be added without impairing the techno-functional properties of the breads. During the first 90 min of proofing, *F. vesiculosus* improved the capacity of doughs to maintain shape by increasing elongational viscosity. However, increased viscosity reduced bubble growth, limiting dough expansion; this impaired the desired final dough porosity. After baking, this increased bread density and apparent crumb modulus. Up to 2% seaweed could be added without altering density, and up to 4% could be added without negatively affecting the acceptability of the density and crumb texture. Additions greater than 4% significantly modified texture parameters of the bread crumb and created a noticeable green colour.

Cox & Abu-Ghannam (2013) significantly enhanced the phytochemical content of wholemeal and white wheat flour breadsticks by adding *Himanthalia elongata*. Seaweed was air-dried, milled, and added to the dough mix in proportions of 5-15%, along with white flour from 10-30% of the overall flour concentration. The remainder was brought to 100% with wholemeal flour. The same amounts of yeast (2.13%), water (34.65%), butter (1.21%) and salt (1.21%) were added to the control and enriched doughs. Doughs were kneaded, proofed (33 °C 45 min), rolled into sticks, proofed again (33 °C 40 min), and baked (210 °C, 20 min). Maximum nutritional enhancement and acceptability of texture and colour was achieved with 17.07% *H. elongata* and 21.89% white flour. Total phenolic content rose by 427%, from 27.67 mg GAE/100 g in the control to 145.88 mg GAE per 100 g in the *H. elongata* enriched sample. Antioxidant capacity, measured by DPPH activity, increased by 87%. Total dietary fibre increased by 71%. Sensory analysis found

that taste, texture, colour, aroma, and overall acceptability of the seaweed-enriched breadsticks was slightly below the control, but not statistically different. The study showed that there is definite potential for the nutritional enrichment of baked goods with seaweed as healthy alternatives to commonly available, nutrient-poor products.

Three seaweeds from Chile, were incorporated into a range of plant and meat-based foods by Astorga-España *et al.* (2017). After drying, the seaweeds were used as ingredients in dishes commonly consumed in the region. *Durvillaea antarctica* was incorporated at 3% in fettuccine; 5% in bread; and 5% in hamburger. *Macrocystis pyrifera* was added at 8% in breadsticks and 7% in fritters; while 28% of *Pyropia columbina* was used in parsley pesto. In the seaweed-enriched dishes, kilocalories per 100 g and protein percentage ranged from: fritters (143 kcal, 6.9%), hamburger (145 kcal, 12.73%), fettuccine (209 kcal, 11.4%), parsley pesto (225 kcal, 4.16%), bread (252 kcal, 9.9%), and breadsticks (316 kcal, 9.5%). Of the three seaweeds, *D. antarctica* exhibited the best protein increase. Compared to regular versions of each dish, fibre contents (dry mass) were significantly greater in seaweed versions, with the minor exception of the breadsticks with 3.6% fibre versus 4.0% in regular breadsticks; and 1.8% fibre in the parsley pesto versus 1.9% in a non-seaweed version. The polyunsaturated fatty acid (PUFA) profiles were improved with seaweed addition. The total lipid content of the seaweed-enriched dishes ranged from 4.1% in *M. pyrifera* breadsticks to 23.1% in *P. columbina* pesto. Of this total lipid percentage, PUFAs were the most abundant fatty acid with the exception of the *D. antarctica* fettuccine in which monounsaturated fatty acids (oleic and nervonic) were more prevalent. Two PUFAs were quantified: linoleic (*n*-6) and α -linolenic (*n*-3) acids. Linoleic acid as a percentage of total lipids in the seaweed-enriched dishes ranged from 22.3% (fettuccine), 32.0% (hamburger), 33.6% (pesto), 37.2% (fritters), 44.5% (bread) to 46.2% (breadsticks). As a percentage of total lipids, α -

linolenic acid was 1.2% (fettuccine), 2.8% (hamburger), 4.3% (fritters), 5.4% (bread), 5.6% (breadsticks), while no α -linolenic acid was detected in the pesto. The total PUFA content (linoleic and α -linolenic combined) of the seaweed-enriched dishes ranged from 23.4% in fettuccine to 51.8% in breadsticks. This is ten to fifty times more than the total PUFA content of the dishes before seaweed addition: 0% (pesto), 0.4% (fettuccine), 1.5% (fritters), 1.6% (breadsticks), 2.1% (bread) and 2.3% (hamburger). The addition of seaweed therefore made a highly significant improvement in essential polyunsaturated fatty acid content.

Other functional nutrients derived from seaweed include polyols, or sugar alcohols, such as mannitol from brown seaweeds (a sweetener), with some reported medicinal properties in the case of reducing oedema and aiding kidney function (Liu *et al.* 2012). Mannitol has a far lower glycaemic index and 40% fewer calories than sucrose and is non-cariogenic (Grembecka 2015). Ulvans are sulphated acidic polysaccharides that occur in green seaweeds and have gelling and rheological properties similar to gum arabic due to solubility in water. Ulvans are being developed as vegan alternatives to beef-derived gelatin due to consumer concerns about bovine BSE contamination (Kraan 2012).

Aside from meat and grain-based foods, seaweed has been used to enhance many other products such dairy, fish, desserts, mayonnaise, sauces and fermented products (Roohinejad *et al.* 2017; Dolea *et al.* 2018; Honold *et al.* 2016; Uchida *et al.* 2018; Abu-Ghannam & Shannon 2017; Pongpichaiudom & Songsermpong 2018). Seaweed as an ingredient has become more acceptable and mainstream outside of Asia. For example, the global furniture company, Ikea, has developed sustainable alternatives to beef meatballs and pork hotdogs using seaweed, vegetables, and insect protein; these were launched in their store restaurants in 2018 (Algaeworld 2018).

The world population is predicted to increase to nine billion by 2050 (Zhou *et al.* 2018). Traditional crop-growing and animal-grazing practices exacerbate competition for land use. Livestock farming is also a major contributor to greenhouse gas emissions of carbon dioxide and methane. Seaweed reduce carbon emissions generated by animal protein production. Seaweeds are suitable for vegan, kosher, and halal diets. As functional foods, seaweeds offer a low-cost, alternative, sustainable source of protein without the saturated fat associated with meat, and are an excellent source of fibre, vitamins, minerals, and polyunsaturated fats.

Epidemiological evidence and dietary intervention studies

The incidence of chronic dietary-associated disorders such as type 2 diabetes, obesity and cardiovascular disease has become a health epidemic in the Western world. First-line therapy by the medical profession often involves pharmaceutical drugs to treat disorders and their symptoms. While these can be helpful, many orthodox drugs have serious side-effects and can lose their efficacy over time. Nutritional intervention has long been proposed as a solution, both to prevent and to alleviate chronic dietary-associated health disorders. However, this is often viewed as anecdotal evidence compared to data from pharmaceutical studies, where effective doses are specified at micro-molar level. The field of natural products chemistry and pharmacognosy has developed in recent years, elucidating numerous plant, fungal and algal compounds with therapeutic effects. Seaweed have unique chemical properties compared to terrestrial plants due to their mineral-rich marine habitat, and the requirements to survive environment. Seaweeds generate antioxidising, antimicrobial and other bioactive agents to combat abiotic stress such as UV photo-damage, high salinity, constant oxygen exposure, and biotic stress from bacterial colonisation and marine herbivores. Various seaweeds as part of the diet, or in the form of isolated extracts, have been documented for disease-preventative effects. Epidemiological evidence from broad

population studies, and from controlled experimental intervention showing the positive effect of seaweed in the diet with respect to numerous human health disorders is discussed.

CANCER: The U.S. Food and Drug Administration, European Pharmacopoeia, and European Food Safety Authority consider seaweeds as food, not medicine (U.S.F.D.A. 2017; Turck *et al.* 2016). However, in many Asian countries, seaweeds are considered as medicinal drugs with details of their effects and directions for use. Examples include the Japanese and Korean pharmacopoeias and the *Chinese Marine Materia Medica* (C.M.M.M.). In traditional Chinese medicine, seaweeds and other marine organisms are documented separately from terrestrial *Materia Medica*. In the C.M.M.M., 171 species of medicinal algae are listed (Fu *et al.* 2016; Qin 2018). Since seaweed has been used as a regular part of the diet and accepted as medicine for millennia in Asia, this may account for the majority of epidemiological evidence being based there (Kim 2017).

A direct relationship has been identified between high levels of seaweed consumption and lower instances of dietary-related disease such as cancer (Iso 2011). At the National Cancer Centre of South Korea, Park *et al.* (2016) assessed the dietary patterns of 923 men and women with an average age of 56 who had previously undergone surgery for colorectal cancer (plus 1846 control participants). Three dietary types were identified: Prudent, Traditional Korean, and Westernised. A highly significant reduction of risk factors for colorectal cancer was identified in subjects in the Prudent group who consumed the most seaweed and vegetables, followed by the Traditional diet (slightly less seaweed), with the highest risk found in the Westernised diet group who consumed little or no seaweed but high levels of red meat and processed foods.

Nelson *et al.* (2017) found the same association in a study of 627 people (age 35–74) across forty-two hospitals in China. Risk factors for the development of biliary tract cancer were

measured using thirty-nine food groups. Only four food groups had either a significantly positive or negative association with risk factors for biliary tract cancer. These groups were seaweed, allium (onions and garlic), salted meats, and preserved vegetables (pickled with salt-brine). The seaweed and allium groups both had an inverse association (i.e. reduced risk) with cancer. The salted meat and preserved vegetable groups had a positive association with cancer (increased risk). The authors surmised that the high fibre and anti-inflammatory properties of seaweed reduced this group's risk of developing cancer.

In a case-control study of breast cancer in South Korea, Yang *et al.* (2010) reported the daily intake of several species of *Pyropia* (as *Porphyra*), to be inversely associated with the risk of developing cancer. *Pyropia* (Gim in Korea) is commonly eaten dried and roasted (Jung & Choe 2017; Herath *et al.* 2018). A cohort of 362 women with histologically confirmed breast cancer (aged 30-65), and 362 control participants of equal age and menopausal status, completed a food frequency questionnaire for twelve months containing 121 food items, including seaweed. An inverse dose-response association with the risk of breast cancer was seen in women who consumed the most Gim over the twelve months (at least 1.17 g day⁻¹, dry mass).

Within the Japanese population of 127 million, where the daily intake of seaweed averages 14.3 g per adult per day (fresh mass), cancer rates are among the lowest in the world (Délérís *et al.* 2016). Fukuda *et al.* (2006) conducted a dietary survey evaluating the dietary fibre intake of 577 Japanese men and women. Amongst eleven fibre-rich foods – including pulses, vegetables, mushrooms, and grains – seaweed contributed the most dietary fibre. In a Japanese Public Health Centre-based Prospective Study for colorectal cancer risk in middle-aged adults, the dietary patterns of 93,062 men and women were assessed over four years. The lowest risk of developing colorectal cancer was associated with the highest seaweed consumption.

Cancer risk increases due to the effect of dietary changes in people who emigrate from Asian to Western countries (Vogel 2018; Deapen *et al.* 2002; Shimizu *et al.* 1991). For example, a population-based, case-control study of breast cancer conducted over four years among Japanese, Chinese, and Filipino women who had emigrated to the U.S.A. (597 subjects, age 20-55, plus 966 controls) showed a six-fold increase in breast cancer risk by migration patterns. Asian-American women born in the U.S.A. also had a cancer risk 60% greater than those born in Asia, while those living in the West for ten years or more had a risk 80% higher than more recent migrants (Ziegler *et al.* 1993). Numerous epidemiological studies of breast cancer have shown that weight is a key risk element. Thus diet, which significantly determines weight, must be considered a fundamental risk factor.

The multiple mechanisms by which seaweeds induce apoptosis in cancer cells has been linked to several compounds such as fucoxanthin, polyphenols and other antioxidants; phlorotannins; iodine; and sulphated polysaccharides such as fucoidan (Jiang & Shi 2018; Gutiérrez-Rodríguez *et al.* 2018; Namvar *et al.* 2012).

Obesity and metabolic disorder

The Organisation for Economic Co-operation and Development reported that in 2015 38.2% of the adult population in the U.S.A. was obese, while only 3.7% in Japan, and 5.3% in Korea (OECD 2017). Obesity – combined with mineral and vitamin deficiency at the same time – has become a health epidemic in many developed regions. Obesity increases the risk of developing other diseases, including type 2 diabetes, hypertension, dyslipidaemia, and coronary heart disease (Medina-Remón *et al.* 2018). Consumption of fibre-rich foods like seaweed, and seaweed isolates including carotenoids and alginate, has been linked to positive effects on appetite, satiety, blood glucose and cholesterol (Brown *et al.* 2014; Kim *et al.* 2008).

Fucoxanthin occurs in brown seaweeds and many microalgae, and has anti-obesity properties (Shannon & Abu-Ghannam 2017). Hitoe & Shimoda (2017) examined the effect of fucoxanthin on fifty women and men (aged 20-59) with a body mass index (BMI) above 26-30 kg/m², and a waist circumference of ≥ 90 cm (women) and ≥ 85 cm (men). Aside from having a BMI above the recommended 18-25 kg/m², subjects were healthy. Either fucoxanthin capsules, or placebos, were given at a dosage of 1 mg or 3 mg/day for four weeks. In the 3 mg/day group, significant decreases occurred in BMI, visceral fat, basal metabolic rate, abdominal fat and circumferences of the neck, arm and thigh compared to the placebo group. In the 1 mg/day group total fat mass, subcutaneous fat area, waist circumference and right thigh circumference were significantly decreased. No adverse effects on blood pressure, pulse rate, blood parameters, or urinalysis parameters were reported during the study.

Abidov *et al.* (2010) administered fucoxanthin capsules over a sixteen-week period to 115 obese, non-diabetic, premenopausal women with a liver fat content above 11%. Each daily dose consisted of a capsule containing 300 mg of dried, brown seaweed extract with a fucoxanthin content of 2.4 mg, combined with 300 mg pomegranate seed oil. The placebo group received olive oil capsules. A significant average weight loss of 4.9 kg, and an increase in resting energy expenditure was observed across the treatment group.

In combination with lifestyle changes such as exercise and dietary changes, seaweed and its extracts could be used as an aid in obesity treatment.

TYPE 2 DIABETES: Worldwide, 108 million adults had diabetes in 1980 (4.7% of global population). This increased to 422 million (8.5% of global population) in 2014. Of these diabetic cases, 90-95% were type 2 diabetes (NCD-RisC 2016). Currently in the United Kingdom, 7.7% of

adults have been diagnosed with type 2 diabetes. In the U.S.A. it is 9.1%, while a further 86 million (26.4% of population) have pre-diabetes (W.H.O. 2016a; W.H.O. 2016b; C.D.C. 2017). Type 2 diabetes mellitus, also known as adult onset diabetes, is a diet-related metabolic disorder that responds well to dietary intervention, unlike the often genetically inherited type 1 diabetes (an autoimmune disorder), that destroys the beta cells of the pancreas, and must be treated with insulin injections. Drugs commonly used to treat type 2 diabetes, such as the α -amylase and α -glucosidase inhibitor Acarbose, may cause gastric discomfort and diarrhoea (Moore 2018).

The *in vivo* anti-diabetic efficacy of seaweed has been proven in many animal studies (Roy *et al.* 2011; Gabbia *et al.* 2017; Zhao *et al.* 2018; Song *et al.* 2018). In human clinical trials, daily supplementation with *Undaria pinnatifida* and *Sacchariza polyschides* (*Gigantea bulbosa*) balances blood glucose levels, decreases serum triglyceride concentrations and increases high-density lipoprotein cholesterol in subjects with type 2 diabetes (Kim *et al.* 2008). Paradis *et al.* (2011) studied the effect of a commercial seaweed extract (*InSea*²) on postprandial plasma glucose and insulin concentrations of twelve women and eleven men (aged 19-59, mean BMI 24.9 kg/m²). The *InSea*² extract used *Fucus vesiculosus* and *Ascophyllum nodosum* and contained natural α -amylase and α -glucosidase inhibitors. The extract had previously demonstrated anti-diabetic properties *in vitro* by inhibiting these digestive enzymes that convert polysaccharides into simple sugars in the intestine and raise blood glucose levels (Roy *et al.* 2011). Thirty minutes before a meal of 50 g white bread, capsules containing 500 mg *InSea*² powder were administered to twelve subjects, while placebo capsules were given to a control group. For 3 hr after meal consumption, plasma glucose levels were reduced by 9% in the seaweed group (1188 mmol l⁻¹ min⁻¹ in the placebo group compared to only 1081 mmol l⁻¹ min⁻¹ in the seaweed group), although the difference was not statistically significant ($P > 0.05$). However, a significant insulin reduction of

12.1% was seen in plasma levels of the seaweed group. In addition, the peripheral insulin sensitivity and muscular glucose uptake (Cederholm index) increased by 7.9%. The improvements in glucose and insulin profiles exerted by the seaweed extracts suggest that they could be used to maintain insulin homeostasis in subjects with type 2 diabetes.

The same *InSea²* extract significantly enhanced postprandial cognitive performance in 33 women and 27 men (age 18-65) who commonly experienced acute postprandial drowsiness (Haskell-Ramsay *et al.* (2018). Attention, performance, and error rates during cognitive tests such as immediate word recall, simple reaction time, digit vigilance, and choice reaction time, were recorded before and after a high-carbohydrate meal. A dose of 500 mg *InSea²* extract was administered 30 min before the carbohydrate (50 g) meal of waffles and pure maple syrup, after which the subjects had to repeat the cognitive tests. The group who consumed the seaweed extract displayed greater accuracy and a 15% reduction in error rates compared to the placebo group. The effects were attributed to modulation of glucose metabolism by the high fibre and polyphenolic contents (20% chlorogenic acid equivalent) of the *InSea²* extract, since similar polyphenols from plants, e.g. cocoa and grapes, increase cerebral blood flow.

Tanemura *et al.* (2014) examined the postprandial blood glucose profiles of 12 healthy adults (eight men, four women, average age 25) after a meal with or without the inclusion of fresh, cooked, whole *U. pinnatifida* (Wakame), or sporophylls of *U. pinnatifida* (Mekabu). An average Japanese cooked breakfast of white rice (200 g), boiled soya beans, broccoli, and potatoes was supplemented with either 70 g fresh Wakame or 70 g fresh Mekabu. The control breakfast contained 455 kcal and 87.5g carbohydrates; while the Wakame breakfast contained 466 kcal and 91.5g carbohydrates; and the Mekabu had 463 kcal and 90 g carbohydrates. Since white rice is a high glycaemic-index food, the study aim was to measure reduction in blood sugar induced by the

seaweed. Half an hour after the meals, glucose levels were significantly lower ($P < 0.05$) in the group who consumed the meal with Mekabu, compared to the control group. The authors surmised that it was the fucoxanthin fraction and the polysaccharide-rich content of the Mekabu sporophylls that exerted the anti-glycaemic effect. The small Mekabu shoots that grow from the parent Wakame seaweed thallus are more viscous when cooked. This is due to their higher content of soluble viscous fibre, which affects carbohydrate metabolism and delays gastric emptying (Tanemura *et al.* 2014). It was concluded that addition of fresh, brown seaweed to meals could be useful in controlling blood glucose levels for people with type 2 diabetes.

The mechanisms of anti-diabetic action by seaweed have been attributed to compounds including phlorotannins, fucoxanthin, polyphenolics, and polysaccharides (Murray *et al.* 2018; Kellogg *et al.* 2014) which inhibit hepatic gluconeogenesis, and reduce the activity of digestive enzymes such as α -amylase, α -glucosidase, lipase and aldose reductase (Sharifuddin *et al.* 2015). Seaweeds and their extracts may alleviate the health epidemic of type 2 diabetes which can lead to nephropathy, blindness, peripheral neuropathy with loss of limb extremities and premature death (Yamazaki *et al.* 2018).

HYPERTENSION AND CARDIAC DISEASE: Hypertension increases the risk of cardiovascular disease to a greater extent than other causes such as cigarette smoking and type 2 diabetes (Roth *et al.* 2015). The aetiology of hypertension and cardiovascular disease involves non-modifiable risk factors such as age, gender, ethnicity and genetics; however, modifiable risk factors such as diet and lifestyle can be improved to greatly reduce the risk of developing high blood pressure and heart disease. Worldwide, heart disease is the leading cause of mortality. In the U.S.A., the annual cost of health care, medication and lost productivity because of heart disease is estimated to be at

least \$200 billion (C.D.C. 2016). Worldwide, 40% of adults aged 25 and over have been diagnosed with high blood pressure. This figure is estimated to increase by 2025 (W.H.O. 2017). Numerous pharmaceuticals for treatment of high blood pressure and heart disease have been developed, such as the angiotensin-I converting enzyme (ACE) inhibitor, Captopril; and the aldosterone inhibitor, Eplerenone. They are effective but may have adverse effects: persistent dry cough, impaired kidney function, and extremely low blood pressure. Statins, such as Lipitor, lower LDL cholesterol to combat hyperlipidaemia and prevent the progress of atherosclerosis. However, statins can have side-effects such as muscle myopathy, elevated blood sugar, impaired liver function, and memory loss (Thompson *et al.* 2016).

An inverse relationship between regular seaweed consumption and reduced risk of hypertension and cardiovascular disease was observed in epidemiological studies (Nanri *et al.* 2017; Maruyama *et al.* 2013; Chu *et al.* 2015; review by Cornish *et al.* 2015). Japan and South Korea have two of the longest average life spans and take less medication for hypertension and cardiovascular disease than populations with lower seaweed intakes (Yamori *et al.* 2017; Lee *et al.* 2016). For example, a fifteen year study of dietary patterns in 79,594 Japanese men and women (aged 45-75) found that a closer adherence to Japanese dietary guidelines was associated with a lower risk of death from all causes and cardiovascular disease, particularly cerebrovascular disease within the experimental population (Kurotani *et al.* 2016).

Hata *et al.* (2001) conducted a dietary intervention trial on thirty-six elderly out-patients with hypertension in Japan, using 3.3 g day⁻¹ of *U. pinnatifida* powder. A control group of eighteen gender-matched subjects (\pm 2 years) were given a placebo. Systolic blood pressure fell significantly in the seaweed group by 13 mmHg after four weeks, and by 8 mmHg after eight weeks. Diastolic pressure fell by 9 mmHg after four weeks, and by 8 mmHg after eight weeks. In

addition, hypercholesterolemia in the treatment group decreased by 8% after four weeks. Teas *et al.* (2009) conducted a similar study with *U. pinnatifida*. Powder (6 g day⁻¹) was administered for one month to 30 men and women with diagnosed metabolic syndrome. In the seaweed group there was a significant decrease in systolic blood pressure (-10.5 mmHg) in all participants, and a reduction in waist circumference (-3.6 cm) (in women only), compared to the placebo group.

Peptide extracts of seaweeds in human cell culture, and animal *in vivo* trials, can significantly reduce blood pressure in single doses and long term administration (Suetsuna & Nakano 2000; Sato *et al.* 2002; Ko *et al.* 2017). Peptides can bind to the active site of the angiotensin-I converting enzyme, thereby reducing blood pressure. For example, peptides extracted from *Gracilariopsis lemaneiformis* had potent ACE-inhibitory activity *in vitro* (Cao *et al.* 2017). Compounds such as sulphated polysaccharides, from *Ulva prolifera* (as *Enteromorpha*) reduce hyperlipidaemia in animal models – one of the principle causes of heart disease (Ren *et al.* 2018; Ren *et al.* 2017). Eicosapentaenoic and docosahexaenoic acids in seaweed may also play a part in the mechanism of heart disease risk reduction. Within cardiac cell membrane phospholipids, these *n*-3 fatty acids can have an antiarrhythmic effect by influencing ionic channels and maintaining intracellular calcium homeostasis (Judé *et al.* 2006; Kanoh *et al.* 2017).

ANTIOXIDANTS: Damage to living cells by free radicals, such as reactive oxygen species, is linked to many chronic disorders. Although the endogenous defence mechanisms of humans can combat oxidation by free radicals to a certain extent, an imbalance still exists in many individuals, especially where the diet is low in antioxidants, and there are high levels of stress. Epidemiological studies have found that dietary compounds such as phlorotannins and fucoxanthin, can reduce the risk of developing disorders associated with free radical cellular damage, including metabolic

syndrome, cancer, cardiovascular disease, osteoporosis, renal disease, Parkinson's, Alzheimer's and neurodegenerative disorders (Cornish & Garbary 2010; Shannon *et al.* 2018; Valko *et al.* 2016; Barbosa *et al.* 2014). The environmental stresses of UV light and oxygen exposure imposed on seaweed in their marine habitat have induced the production of antioxidant metabolites. Numerous studies have reported that the clinical bioactivity of algal extracts is directly related to their antioxidant capacity as measured by assays such as DPPH and ORAC (Narasimhan *et al.* 2013; Pinteus *et al.* 2017a)

Kang *et al.* (2012) studied *Saccharina japonica* (as *Laminaria*), for its antioxidant health potential. This species has been used as a folk remedy in Korea for centuries. A human clinical trial examined whether *S. japonica* could enhance the antioxidant defence system of 48 Korean men (age 25-60). The alga was fermented with a lactic acid producing bacterium, *Lactobacillus brevis*, for five days to potentially augment the seaweed's bioactivity and digestibility. The fermented seaweed was dried, and 250 mg of the powder was encapsulated. For one month, six capsules per day (1.5 g of alga) were administered to healthy subjects, but who had high levels (< 132 U/L) of the enzyme, γ -glutamyltransferase (γ -GT). Volunteers with high levels of γ -GT were selected for the study because this enzyme is a predictive biomarker of cellular antioxidant inadequacy, disease risk, and is linked to underlying alcohol-related liver dysfunction (Koenig & Seneff 2015). Serum γ -GT levels decreased significantly from 102.21 to 78.70 U/L in the seaweed group after one month, and actually increased from 104.25 to 116.75 U/L in the placebo group. The blood serum levels of four other biological compounds were measured: malondialdehyde, catalase, superoxide dismutase, and glutathione peroxidase, chosen because they are involved in the cell defences against free radical damage by eliminating free radicals and reactive oxygen species. The aim was to increase blood serum levels of these enzymes. Malondialdehyde is a

product of lipid peroxidation in the body; therefore, its reduction is an indication of a healthy antioxidant defence system. Administering fermented *S. japonica* for one month significantly increased levels of catalase and superoxide dismutase; while levels of γ -GT and malondialdehyde were significantly decreased compared to the placebo group. No difference was observed in glutathione peroxidase levels between the treatment and control groups. These findings are very encouraging since healthy biomarkers of the antioxidant defence system were increased, while unhealthy biomarkers were simultaneously decreased. The authors concluded that fermented *S. japonica* could be developed as an antioxidant functional food ingredient.

Phlorotannins are compounds unique to brown seaweeds. They are similar to polyphenols, but are composed of repeating phloroglucinol units (three hydroxyls bound to a benzene ring) rather than phenolic units (one hydroxyl bound to benzene; Sathya *et al.* 2017). Phlorotannins are powerful antioxidants due to their ability to act as chelating agents with reactive oxygen species, thereby preventing oxidative stress and cell damage. Pinteus *et al.* (2017b) evaluated the protective effect of phlorotannin extracts on human (MCF-7) cells against oxidative stress using a methanolic extraction of *Fucus spiralis*. Oxidative stress was induced in cells by addition of hydrogen peroxide. Cell death resulting from oxidative stress decreased 51% with the addition of phlorotannin extracts (1 mg m^{-1} for 24 h), compared to the control. It was concluded that the phlorotannin extracts maintained normal membrane potential in the cells by decreasing caspase-9 activity which is involved in cell death mechanisms, thereby protecting the cells from oxidative damage.

A human clinical trial by Shin *et al.* (2012) found that phlorotannins extracted from *Ecklonia cava* had an antioxidant effect, by alleviating hyperglycaemia-induced oxidative stress. Ninety-seven overweight men and women (average age 40, BMI 26.5 kg/m^2) were given either

144 mg day⁻¹ or 72 mg day⁻¹ of the phlorotannin extract for three months, while a parallel control group received a placebo. Both dosage groups experienced a significant reduction of total cholesterol, LDL cholesterol, BMI, atherogenic index, and waist circumference compared to a placebo group. The higher dosage group also experienced a significant decrease in blood glucose and systolic blood pressure, and increased HDL cholesterol. Lee & Jeon (2015) also isolated antioxidant-rich phlorotannins from *E. cava* and administered 1500 mg day⁻¹ of the extract to 80 pre-diabetic adults (aged 20-65). After three months, a significant decrease in postprandial glucose levels was measured in the phlorotannin extract group. The authors concluded that the phlorotannin-rich (46%) extract, and the high content of the phlorotannin dieckol (10%) that was responsible for the anti-diabetic effect.

Fucoxanthin is a tetraterpenoid, carotenoid reported to be a powerful dietary antioxidant. Due to its unusual chemical structure, fucoxanthin donates an electron to quench reactive oxygen species, instead of a proton, as in most antioxidants such as β -carotene or ascorbic acid. Fucoxanthin can also function as an antioxidant under anoxic conditions, whereas most antioxidants cannot. This is advantageous for humans since there are low levels of oxygen within physiological tissues (Yan *et al.* 1999; D'Orazio *et al.* 2012; Abu-Ghannam & Shannon 2017). Jang *et al.* (2018) demonstrated the antioxidative protective effect of fucoxanthin extracted from *L. japonica* on human liver (HepG2) cells *in vitro*. Oxidative stress was induced in the HepG2 cells by the addition of arachidonic acid and iron. Cells pre-treated with fucoxanthin (30 μ g ml⁻¹ for 1 hr) had significantly greater viability than untreated cells. The mechanism by which fucoxanthin protected liver cells against oxidative stress was thought to be associated with the LKB1-AMPK α signalling pathway. LKB1 is a liver kinase. AMPK α is adenosine monophosphate-activated protein kinase – an enzyme involved in energy homeostasis. Jang *et al.* (2018) concluded

that the LKB1-AMPK α signalling pathway was enhanced, which triggered autophagy, evidenced by the significant increase of two autophagic protein markers (beclin-1 and LC3II). The enhanced autophagy helped maintain homeostasis and survival in the HepG2 cells treated with fucoxanthin. *Ex vivo* studies in animal models have reported that dietary supplementation of fucoxanthin (0.2% of total food for one month) improved the antioxidant capacity of blood serum levels in obese rats via activation of the nuclear erythroid factor like-2 pathway, which regulates antioxidative defence responses and genes such as NQO1. NAD(P)H quinone oxidoreductase (NQO1) is one of the genes involved in the maintenance of intracellular reducing-potential, and the scavenging of reactive oxygen or nitrogen species (Ha *et al.* 2013). Zaragozá *et al.* (2008) used ethanol-extraction to isolate fucoxanthin from *F. vesiculosus*. For four weeks, rats were fed fucoxanthin as part of their food (0.2 g kg⁻¹ body weight). Significantly increased antioxidant activity was observed in *ex vivo* assays of erythrocytes and plasma. Activity of copper-zinc superoxide dismutase (Cu-Zn SOD) was measured. Cu-Zn SOD is involved in the protective mechanism against oxidative stress in cells. Cu-Zn SOD activity increased 32% in rats fed with fucoxanthin after four weeks.

ANTIBACTERIAL: The global burden of infectious disease caused by bacteria, viruses, fungi, and protozoans continues to grow despite the development of antibiotics in the 1940's. In the Western world, the issue is not availability of antimicrobial treatments, but the developed immunity of micro-organisms to pharmaceutical drugs and disinfectants. Bacteria, in particular, pose a threat due to their ability to evolve and survive within human hosts and on surfaces. Currently, at least 700,000 people die worldwide each year from drug-resistant bacterial infections such as *Pseudomonas aeruginosa*, *Helicobacter pylori*, *Mycobacterium tuberculosis*, *Neisseria gonorrhoeae*, *Haemophilus influenzae*, and methicillin-resistant *Staphylococcus aureus* (MRSA).

It is estimated that by 2050, drug-resistant bacteria will cause 10 million deaths annually worldwide – more than cancer (Willyard 2017; Richter & Hergenrother 2018). In the fight against resistant infectious bacteria, seaweed products offer some alternatives to orthodox antibiotics.

Several compounds that occur naturally in seaweed have demonstrated antibacterial efficacy equal to that of pharmaceutical drugs. These include phlorotannins, polysaccharides, fatty acids, terpenes, peptides, lactones, pigments, and chrysophaeantins (Shannon & Abu-Ghannam 2016). The assessment of antimicrobial activity in human clinical trials is very restricted, therefore *in vitro*, *ex vivo*, and animal studies are key to assessing novel algal antimicrobial extracts. Biofilm formation by infectious bacteria is one of the most problematic issues affecting antibiotic treatment, medical devices, implants, and surface contamination. Many species of bacteria secrete a combination of polysaccharides, proteins, and extracellular DNA that form a protective layer under which they can thrive. It is estimated that biofilms are present in 65-85% of all bacterial infections (Aynapudi *et al.* 2017). These films can form in wounds and inside lungs. For those suffering from cystic fibrosis, biofilm formation in lungs by *Pseudomonas aeruginosa* can be fatal. Polysaccharides from algae, such as alginates, have been studied for their activity against biofilms. Currently, a biopolymer with the commercial name OligoG derived from *Laminaria hyperborea* is undergoing human clinical trials in cystic fibrosis patients (AlgiPharma 2018). OligoG is an edible alginate oligosaccharide derivative which has already been shown *in vitro* to disrupt *P. aeruginosa* biofilms.

Pritchard *et al.* (2017) used artificial sputum medium with clinical cystic fibrosis isolates to measure the effect of OligoG on a model, multidrug-resistant pseudomonal biofilm, and also evaluated whether OligoG (CF-5/20) could reduce the dose of drugs normally required to treat infections. The antibiotic, Colistin, was used as a pharmaceutical standard in combination with the

seaweed derivative. Treatment with 2% OligoG significantly disrupted colony formation of *P. aeruginosa* by reducing quorum-sensing signalling among the bacterial cells. In a control medium without OligoG, the minimum inhibitory concentration (MIC) of the antibiotic Colistin increased from 0.1 to 0.4 $\mu\text{g ml}^{-1}$ in the presence of the multidrug-resistant *P. aeruginosa*. Therefore, without the seaweed extract, four times more antibiotic would have to be administered to patients. However, the addition of 2% seaweed extract led to Colistin retaining its antimicrobial activity at the lower dose.

The same extract of *L. hyperborea* successfully treated *P. aeruginosa* infection in mice (Hengzhuang *et al.* 2016). Lung biofilms were established in mice by tracheal instillation of *P. aeruginosa* (1×10^8 CFU ml^{-1}) on alginate beads; treatment groups received the same alginate beads combined with saline and a dose of 0.2%, 1%, or 5% OligoG (CF-5/20). Within 24 h, a highly significant 2.5 log reduction bacterial colony forming units was observed in the lungs of the 5% OligoG group. Blood tests revealed that levels of the interleukin, IL-1 α , also decreased in the treated mice – proof of a general reduction in inflammatory response. In addition to the murine study, Hengzhuang *et al.* (2016) conducted *in vitro* biofilm plate tests on two strains of *P. aeruginosa* (PAO1 and NH57388A) using OligoG combined with Colistin and another antibiotic Ciprofloxacin. Minimum biofilm eradication concentration (MBEC) assays were carried out on the plated bacteria and antibiotic/OligoG combinations. A synergistic effect was observed for both antibiotics. However, the most significant was a 128-fold reduction in the amount of Colistin required to treat the NH57388A strain of *P. aeruginosa*. After 8 h, the 5% OligoG sample had reduced the Colistin MBEC from 512 $\mu\text{g ml}^{-1}$ to just 4 $\mu\text{g ml}^{-1}$. There also was synergism between Ciprofloxacin and OligoG for the PAO1 *P. aeruginosa* strain, but not for NH58388A. Even at concentrations of 1% OligoG, synergistic effects were observed, with reductions in the MBECs

for both drugs. The EC₅₀ for OligoG was calculated to be 0.8%. Sigmoidal modelling predicted that one dose of 3% OligoG would eradicate of 99% of the biofilm infection within 24 h.

These studies highlight the potential of some seaweed derivatives to treat multidrug-resistant biofilm infections, while simultaneously dramatically reducing the dosage of antibiotic required. The current, on-going human clinical trial with OligoG is investigating whether a fine-powdered form of alginate can be inhaled by people suffering from cystic fibrosis to combat lung biofilm formation (AlgiPharma 2018).

Other seaweed extracts have shown *in vitro* antibacterial efficacy. For example, phlorotannins inhibit oxidative phosphorylation in bacteria, and bind with proteins such as enzymes and cell membrane structures, resulting in lysis of the cell. Wei *et al.* (2015) inactivated *Vibrio parahaemolyticus* with phlorotannins from *Sargassum thunbergii* with a potential to develop the extracts as food antimicrobials, and aquacultured drugs. The fatty acids, cyclopentaneacetic acid, and 10,13-octadecadienoic acid, were isolated from *Sargassum vulgare* and *S. fusiforme* by El Shafay *et al.* (2016). The cell walls of *Staphylococcus aureus* and *Klebsiella pneumoniae* were visibly perforated, killing the bacteria, after treatment with of *S. fusiforme* (100 µl diethyl ether extract) and *S. vulgare* (50 µl ethanolic extract). Polysaccharides, such as fucoidans, have been used as antibacterial agents. (Huang *et al.* (2018)) extracted fucoidan from *Sargassum crassifolium* using hot water and degradation by hydrogen peroxide and ascorbic acid. The extract had *in vitro* antibacterial activity against Gram-positive *S. aureus* and Gram-negative *E. coli*. The authors surmised that this was due to the ability of the negatively charged sulphated fucoidan to trap cationic molecules. This meant that positively charged molecules, such as calcium and other minerals in the culture medium, were made unavailable to the bacteria, thereby starving them. The fucoidan also reversed H₂O₂-induced cell death in 3T3-L1 adipocytes. It was concluded

that the edible seaweed extract had potential as a natural antibacterial and anti-adipogenic agent for functional food and cosmetic applications (review by Jahan *et al.* 2017).

The emerging field of medical textiles provides opportunities for seaweed for its wound healing properties in combination with other natural fibres to make absorbent, biodegradable, dressings that minimise bacterial contamination. Janarthanan & Senthil Kumar (2018) developed a bioactive wound dressing material using filamentous strands of *Chaetomorpha linum* woven with cotton in the ratio of 70:30 seaweed:cotton. Sections of the dressing were soaked in three pathogenic bacterial solutions and tested for bacterial inhibitory power after 12 h. *E. coli* was inhibited by 95%; *S. aureus* by 90%; and *P. aeruginosa* by 88%. The seaweed gauze dressing was also more absorbent than the 100% cotton version, making it more effective for absorbing wound exudate. The bioactive gauze fabric was recommended for use as a non-implantable material in bandages, surgical masks and gowns, or as a multi-use hygienic textile.

Lokhande *et al.* (2018) used κ -carrageenan (κ CA), to produce a haemostatic product, i.e. an agent that stops bleeding. However, rather than a surface wound dressing, the nano-engineered agent controls internal bleeding and is injected as a liquid. The formulation consisted of 1% κ CA and 2% nanosilicate [Na^+ , Mg^{2+} , $\text{Si}(\text{OH})_4$, Li^+]. A two-fold improvement in clotting time and wound healing was demonstrated *in vitro*. The authors concluded that combining nanosilicates with κ CA increased protein adsorption on nanocomposite hydrogels, enhanced cell adhesion and spreading, and increased platelet-binding, thereby reducing blood-clotting time. Since haemorrhage from internal bleeding is the main cause of death in battlefield wounds, the hydrogel could be used as an injectable haemostat in emergency situations or during surgery.

CONCLUSIONS

Seaweeds are a sustainable source of bioactive compounds for human health and functional food applications. The global burden of non-communicable, lifestyle-related diseases such as type 2 diabetes, hypertension, obesity, cancer, antibiotic resistance, and heart disease place a huge strain on the finances and resources of health services in affected countries. This may be alleviated by the inclusion of seaweed and seaweed isolates in the diet, as part of overall lifestyle improvement. Prescription drugs have many unpleasant and often serious side effects, whereas none have been reported from seaweed dietary therapy or extracts. Apart from side-effects, orthodox drugs are also a financial burden on patients. The global seaweed market was valued at \$10.4 billion in 2015 and is projected to reach \$14.7 billion by 2021. It is driven mainly by the growing use of seaweed for medicinal applications. ‘Food as pharma’ could be promoted in terms of the natural health and nutritional benefits of dietary macroalgae based on epidemiological studies of long-term impact, and intervention studies - the ‘dose concept’ of part of your ‘five a day’. Having reviewed the literature on the benefits of seaweed consumption, data from scientific medical studies may inform public health systems in the design of dietary intervention plans, and may be beneficial for policy makers, educators, practitioners, researchers and academics who contribute to promotion of public health. Collaboration in research and public programs is needed to prevent disease through integration in society at all levels. The techno-functional properties of seaweed can be incorporated into food: from fat replacers, to antioxidant, fibre, and antimicrobial enhancement. Seaweeds are a low-calorie food that add protein without saturated fat; and are an excellent dietary lipid alternative to EPA and DHA derived from fish. Seaweed is a sustainable, low cost crop that takes up no land space and produces no green-house gases. Farmed seaweed produces more food biomass per acre of ocean than crops do on land. Cultivation processes are important and can be

developed to justify the production of seaweed industrially, instead of its current form as a cottage industry in many countries. A model could be followed similar to that of countries where marine agronomy, mariculture, and bioengineering of seaweed are common practices. The concept of seaweed ‘from farm to fork’ can be developed in terms of the whole cultivation of seaweed, using technologies already available in the meat and dairy industries. Before seaweed can become mainstream in Western countries, culinary issues must be dealt with. Seaweed is not familiar to many populations outside Asia, but can be promoted through clever product design, development, advertising, and incorporation into everyday products already being consumed. Unpalatable odour compounds and textures in seaweed foods can be reduced by clever ingredient formulations, processing, or by fermentation with lactic acid bacteria or *Aspergillus oryzae*. Chemical synthesis of algal bioactive compounds could be developed to increase supply and protect the marine environment from over-harvesting.

REFERENCES

- Abidov M., Ramazanov Z., Seifulla R. & Grachev S. 2010. The effects of Xanthigen™ in the weight management of obese premenopausal women with non-alcoholic fatty liver disease and normal liver fat. *Diabetes, Obesity and Metabolism* 12: 72–81. DOI: 10.1111/j.1463-1326.2009.01132.x.
- Abomohra A.E.-F., El-Naggar A.H. & Baeshen A.A. 2018. Potential of macroalgae for biodiesel production: screening and evaluation studies. *Journal of Bioscience and Bioengineering* 125: 231-237. DOI: 10.1016/j.jbiosc.2017.08.020.
- Abu-Ghannam N. & Shannon E. 2017. Seaweed carotenoid, fucoxanthin, as functional food. In: *Microbial functional foods and nutraceuticals* (Ed. by V. Gupta, H. Treichel, V. Shapaval, L.A. de Oliveira & T. M), pp. 39–64. John Wiley & Sons, Hoboken, New Jersey, USA.

- Algaeworld. 2018. Algae food and feed - bug burgers, algae buns and herb ice cream: Ikea's fast food of the future. <http://news.algaeworld.org/2018/03/bug-burgers-algae-buns-and-herb-ice-cream-ikeas-fast-food-of-the-future/>; searched on 17 February 2019.
- AlgiPharma. 2018. Safety, tolerability and preliminary efficacy of inhaled *OligoG* in subjects with cystic fibrosis. Phase 2. ClinicalTrials.gov Identifier: NCT01465529. <http://algipharma.com/clinical-trials/>; searched on 24 January 2019.
- Arufe S., Della Valle G., Chiron H., Chenlo F., Sineiro J. & Moreira R. 2018. Effect of brown seaweed powder on physical and textural properties of wheat bread. *European Food Research and Technology* 244: 1–10. DOI: 10.1007/s00217-017-2929-8.
- Astorga-España M., Mansilla A., Ojeda J., Marambio J., Rosenfeld S., Mendez F., Provoste J. & Ocaranza P. 2017. Nutritional properties of dishes prepared with sub-Antarctic macroalgae—an opportunity for healthy eating. *Journal of Applied Phycology* 29: 2399–2406. DOI: 10.1007/s10811-017-1131-5.
- Astorga-España M.S., Rodríguez Galdón B., Rodríguez Rodríguez E.M. & Díaz Romero C. 2015. Mineral and trace element concentrations in seaweeds from the sub-Antarctic ecoregion of Magallanes (Chile). *Journal of Food Composition and Analysis* 39: 69–76. DOI: 10.1016/j.jfca.2014.11.010.
- Aynapudi J., El-Rami F., Ge X., Stone V., Zhu B., Kitten T. & Xu P. 2017. Involvement of signal peptidase I in *Streptococcus sanguinis* biofilm formation. *Microbiology* 163: 1306–1318. DOI: 10.1099/mic.0.000516.
- Belattmania Z., Engelen A., Pereira H., Serrao E., Custódio L., Varela J., Zrid R., Reani A. & Sabour B. 2018. Fatty acid composition and nutraceutical perspectives of brown seaweeds from the Atlantic coast of Morocco. *International Food Research Journal* 25: 1520–1527.

B.N.F. 2016. British Nutrition Foundation. Nutrition requirements: reference nutrient intakes for minerals.

https://www.nutrition.org.uk/attachments/article/234/Nutrition%20Requirements_Revision%20Oct%202016.pdf; searched on 11 January 2019.

Barbosa M., Valentão P. & Andrade P.B. 2014. Bioactive compounds from macroalgae in the new millennium: implications for neurodegenerative diseases. *Marine Drugs* 12: 4934–4972. DOI: 10.3390/md12094934.

Biancarosa I., Belghit I., Bruckner C.G., Liland N.S., Waagbø R., Amlund H., Heesch S. & Lock E.J. 2018. Chemical characterization of 21 species of marine macroalgae common in Norwegian waters: benefits of and limitations to their potential use in food and feed. *Journal of the Science of Food and Agriculture* 98: 2035–2042. DOI: 10.1002/jsfa.8798.

Branscheid W. & Judas M. 2011. Detection of bone in meat. In: *Handbook of analysis of edible animal by-products* (Ed. by L. Nollet & F. Toldra), pp. 278. CRC Press, Boca Raton, Florida, USA.

Brown E.M., Allsopp P.J., Magee P.J., Gill C.I., Nitecki S., Strain C.R. & McSorley E.M. 2014. Seaweed and human health. *Nutrition Reviews* 72: 205–216. DOI: 10.1111/nure.12091.

C.D.C. 2016. Centers for Disease Control and Prevention (CDC). National Center for Health Statistics. Heart Disease Fact Sheet, 1999-2015. <https://wonder.cdc.gov/mcd-icd10.html>; searched on 12 January 2019.

C.D.C. 2017. Centers for Disease Control and Prevention (CDC). Chronic disease prevention and health promotion publications 2016–2017. At a glance fact sheets - Diabetes. <https://www.cdc.gov/chronicdisease/resources/publications/aag/diabetes.htm>; searched on 22 January 2019.

- Cao D., Lv X., Xu X., Yu H., Sun X. & Xu N. 2017. Purification and identification of a novel ACE inhibitory peptide from marine alga *Gracilariopsis lemaneiformis* protein hydrolysate. *European Food Research and Technology* 243: 1829–1837. DOI: 10.1007/s00217-017-2886-2.
- Černá M. 2011. Chapter 24 - Seaweed proteins and amino acids as nutraceuticals. In: *Advances in food and nutrition research* (Ed. by S.-K. Kim), pp. 297–312. Academic Press, San Diego, California, USA.
- Chu S.-M., Shih W.-T., Yang Y.-H., Chen P.-C. & Chu Y.-H. 2015. Use of traditional Chinese medicine in patients with hyperlipidemia: a population-based study in Taiwan. *Journal of Ethnopharmacology* 168: 129-135. DOI: 10.1016/j.jep.2015.03.047.
- Chye J.T.T., Jun L.Y., Yon L.S., Pan S. & Danquah M.K. 2018. Biofuel production from algal biomass. In: *Bioenergy and biofuels* (Ed. by O. Konur), pp. 621. CRC Press, Boca Raton, Florida, USA.
- Cofrades S., Benedi J., Garcimartin A., Sánchez-Muniz F. & Jimenez-Colmenero F. 2017. A comprehensive approach to formulation of seaweed-enriched meat products: from technological development to assessment of healthy properties. *Food Research International* 99: 1084–1094. DOI: 10.1016/j.foodres.2016.06.029.
- Cornish M.L., Critchley A.T. & Mouritsen O.G. 2015. A role of dietary macroalgae in the amelioration of certain risk factors associated with cardiovascular disease. *Phycologia* 54: 649–666.
- Cornish M.L. & Garbary D.J. 2010. Antioxidants from macroalgae: potential applications in human health and nutrition. *Algae* 25: 155–171.

- Cox S. & Abu-Ghannam N. 2013. Enhancement of the phytochemical and fibre content of beef patties with *Himanthalia elongata* seaweed. *International Journal of Food Science & Technology* 48: 2239–2249. DOI: 10.1111/ijfs.12210.
- Cox S. & Abu-Ghannam N. 2013. Incorporation of *Himanthalia elongata* seaweed to enhance the phytochemical content of breadsticks using response surface methodology. *International Food Research Journal* 20: 1537–1545. DOI: 10.21427/D7DS5R.
- D’Orazio N., Gemello E., Gammone M.A., de Girolamo M., Ficoneri C. & Riccioni G. 2012. Fucoxanthin: a treasure from the sea. *Marine Drugs* 10: 604–616. DOI: 10.3390/md10030604.
- Davis G.D.J. & Vasanthi A.H.R. 2011. Seaweed metabolite database: A database of natural compounds from marine algae. *Bioinformatics* 5: 361–364. DOI: 10.6026/97320630005361.
- Dawczynski C., Schubert R. & Jahreis G. 2007. Amino acids, fatty acids, and dietary fibre in edible seaweed products. *Food Chemistry* 103: 891–899. DOI: 10.1016/j.foodchem.2006.09.041.
- Deapen D., Liu L., Perkins C., Bernstein L. & Ross R.K. 2002. Rapidly rising breast cancer incidence rates among Asian-American women. *International Journal of Cancer* 99: 747–750. DOI: 10.1002/ijc.10415.
- Déléris P., Nazih H. & Bard J.M. 2016. Chapter 10 - Seaweeds in Human Health. In: *Seaweed in health and disease prevention* (Ed. by J. Fleurence & I. Levine), pp. 319–367. Academic Press, San Diego, California, USA.
- Dillehay T.D., Ramirez C., Pino M., Collins M.B., Rossen J. & Pino-Navarro J. 2008. Monte Verde: seaweed, food, medicine, and the peopling of South America. *Science* 320: 784–786. DOI: 10.1126/science.1156533.

- Dolea D., Rizo A., Fuentes A., Barat J. & Fernández-Segovia I. 2018. Effect of thyme and oregano essential oils on the shelf life of salmon and seaweed burgers. *Food Science and Technology International* 24: 394–403. DOI: 10.1177/1082013218759364.
- El Maghraby D.M. & Fakhry E.M. 2015. Lipid content and fatty acid composition of Mediterranean macroalgae as dynamic factors for biodiesel production. *Oceanologia* 57: 86–92. DOI: 10.1016/j.oceano.2014.08.001.
- El Shafay S.M., Ali S.S. & El-Sheekh M.M. 2016. Antimicrobial activity of some seaweeds species from Red sea, against multidrug resistant bacteria. *The Egyptian Journal of Aquatic Research* 42: 65–74. DOI: 10.1016/j.ejar.2015.11.006.
- Finglas P., Roe M., Pinchen H., Berry R., Church S., Dodhia S., Powel N., Farron-Wilson M., McCardle J. & Swan G. 2015. *McCance and Widdowson's the composition of foods - Food Standard Agency*, ed. 7. Royal Society of Chemistry, Cambridge, UK.
- Fu X.-M., Zhang M.-Q., Shao C.-L., Li G.-Q., Bai H., Dai G.-L., Chen Q.-W., Kong W., Fu X.-J. & Wang C.-Y. 2016. Chinese marine materia medica resources: status and potential. *Marine Drugs* 14: 46. DOI: 10.3390/md14030046.
- Fukuda S., Saito H., Nakaji S., Yamada M., Ebine N., Tsushima E., Oka E., Kumeta K., Tsukamoto T. & Tokunaga S. 2006. Pattern of dietary fiber intake among the Japanese general population. *European Journal of Clinical Nutrition* 61: 99. DOI: 10.1038/sj.ejcn.1602505.
- Gabbia D., Dall'Acqua S., Di Gangi I.M., Bogialli S., Caputi V., Albertoni L., Marsilio I., Paccagnella N., Carrara M., Giron M.C. & De Martin S. 2017. The phytocomplex from *Fucus vesiculosus* and *Ascophyllum nodosum* controls postprandial plasma glucose levels: an *in vitro* and *in vivo* study in a mouse model of NASH. *Marine Drugs* 15: 41. DOI: 10.3390/md15020041.

- Grembecka M. 2015. Sugar alcohols - their role in the modern world of sweeteners: a review. *European Food Research and Technology* 241: 1–14. DOI: 10.1007/s00217-015-2437-7.
- Guiry M.D. & Guiry G.M. 2019. *AlgaeBase*. World-wide electronic publication, National University of Ireland, Galway. <http://www.algaebase.org>; searched 27 June 2019.
- Gutiérrez-Rodríguez A.G., Juárez-Portilla C., Olivares-Bañuelos T. & Zepeda R.C. 2018. Anticancer activity of seaweeds. *Drug Discovery Today* 23: 434–447. DOI: 10.1016/j.drudis.2017.10.019.
- Ha A.W., Na S.J. & Kim W.K. 2013. Antioxidant effects of fucoxanthin rich powder in rats fed with high fat diet. *Nutrition Research and Practice* 7: 475–480. DOI: 10.4162/nrp.2013.7.6.475.
- Haskell-Ramsay C.F., Jackson P.A., Dodd F.L., Forster J.S., Bérubé J., Levinton C. & Kennedy D.O. 2018. Acute post-prandial cognitive effects of brown seaweed extract in humans. *Nutrients* 10: 85. DOI: 10.3390/nu10010085.
- Hata Y., Nakajima K., Uchida J.-I., Hidaka H. & Nakano T. 2001. Clinical effects of brown seaweed, *Undaria pinnatifida* (Wakame), on blood pressure in hypertensive subjects. *Journal of Clinical Biochemistry and Nutrition* 30: 43–53. DOI: 10.3164/jcbtn.30.43.
- Herath K.M., Lee J.H., Cho J., Kim A., Shin S.M., Kim B., Jeon Y.J. & Jee Y. 2018. Immunostimulatory effect of pepsin enzymatic extract from *Porphyra yezoensis* on murine splenocytes. *Journal of the Science of Food and Agriculture* 98: 3400–3408. DOI: 10.1002/jsfa.8851.
- Hengzhuang W., Song Z., Ciofu O., Onsoyen E., Rye P.D. & Hoiby N. 2016. OligoG CF-5/20 disruption of mucoid *Pseudomonas aeruginosa* biofilm in a murine lung infection model. *Antimicrobial Agents and Chemotherapy* 60: 2620–2626. DOI: 10.1128/AAC.01721-15.

- Hitoe S. & Shimoda H. 2017. Seaweed fucoxanthin supplementation improves obesity parameters in mildly obese Japanese subjects. *Functional Foods in Health and Disease* 7: 246–262. DOI: 10.31989/ffhd.v7i4.333.
- Honold P.J., Jacobsen C., Jónsdóttir R., Kristinsson H.G. & Hermund D.B. 2016. Potential seaweed-based food ingredients to inhibit lipid oxidation in fish-oil-enriched mayonnaise. *European Food Research and Technology* 242: 571–584. DOI: 10.1007/s00217-015-2567-y.
- Huang C.-Y., Kuo C.-H. & Lee C.-H. 2018. Antibacterial and antioxidant capacities and attenuation of lipid accumulation in 3T3-L1 adipocytes by low-molecular-weight fucoidans prepared from compressional-puffing-pretreated *Sargassum crassifolium*. *Marine Drugs* 16: 24. DOI: 10.3390/md16010024.
- HUSTED, K.S., & BOUZINOVA, E.V. (2016). The importance of n-6/n-3 fatty acids ratio in the major depressive disorder. *Medicina* 52: 139–147. DOI: 10.1016/j.medici.2016.05.003.
- Iso H. 2011. Lifestyle and cardiovascular disease in Japan. *Journal of Atherosclerosis and Thrombosis* 18: 83–88. DOI: 10.5551/jat.6866.
- Jahan A., Ahmad I.Z., Fatima N., Ansari V.A. & Akhtar J. 2017. Algal bioactive compounds in the cosmeceutical industry: a review. *Phycologia* 56: 410–422. DOI: 10.2216/15.58.1
- Janarthanan M. & Senthil Kumar M. 2018. A modern development of bioactive wound dressing material using *Chaetomorpha linum* seaweed/cotton blended fabric. *International Journal of Clothing Science and Technology* 30: 16–28. DOI: 10.1108/IJCST-12-2016-0142
- Jang E.J., Kim S.C., Lee J.-H., Lee J.R., Kim I.K., Baek S.Y. & Kim Y.W. 2018. Fucoxanthin, the constituent of *Laminaria japonica*, triggers AMPK-mediated cytoprotection and

- autophagy in hepatocytes under oxidative stress. *BMC Complementary and Alternative Medicine* 18: 97. DOI: 10.1186/s12906-018-2164-2.
- Jiang J. & Shi S. 2018. Seaweeds and cancer prevention. In: *Bioactive seaweeds for food applications*. (Ed. by Y. Qin), pp. 269–290. Academic Press, San Diego, California, USA.
- Jiao G., Yu G., Zhang J. & Ewart H.S. 2011. Chemical structures and bioactivities of sulfated polysaccharides from marine algae. *Marine Drugs* 9: 196–223. DOI: 10.3390/md9020196.
- Jimenez-Escrig A. & Goñi C.I. 1999. Nutritional evaluation and physiological effects of edible seaweeds. *Archivos Latinoamericanos de Nutricion* 49: 114–120. <https://www.ncbi.nlm.nih.gov/pubmed/10488389>; searched on 03 January 2019.
- Judé S., Roger S., Martel E., Besson P., Richard S., Bougnoux P., Champeroux P. & Le Guennec J.-Y. 2006. Dietary long-chain omega-3 fatty acids of marine origin: A comparison of their protective effects on coronary heart disease and breast cancers. *Progress in Biophysics and Molecular Biology* 90: 299–325. DOI: 10.1016/j.pbiomolbio.2005.05.006.
- JUNG, L. & CHOE, E. (2017). Lipid oxidation-related characteristics of gim bugak (Korean fried cuisine with *Porphyra*) affected by frying oil. *Food Science and Biotechnology* 26: 623. DOI: [10.1007/s10068-017-0088-0](https://doi.org/10.1007/s10068-017-0088-0).
- Kang Y.M., Lee B.-J., Kim J.I., Nam B.-H., Cha J.-Y., Kim Y.-M., Ahn C.-B., Choi J.-S., Choi I.S. & Je J.-Y. 2012. Antioxidant effects of fermented sea tangle (*Laminaria japonica*) by *Lactobacillus brevis* BJ20 in individuals with high level of γ -GT: A randomized, double-blind, and placebo-controlled clinical study. *Food and Chemical Toxicology* 50: 1166–1169. DOI: 10.1016/j.fct.2011.11.026.

- Kanoh M., Inai K., Shinohara T., Tomimatsu H. & Nakanishi T. 2017. Clinical implications of eicosapentaenoic acid/arachidonic acid ratio (EPA/AA) in adult patients with congenital heart disease. *Heart and Vessels* 32: 1513–1522. DOI: 10.1007/s00380-017-1015-2.
- Kellogg J., Grace M. & Lila M. 2014. Phlorotannins from Alaskan seaweed inhibit carbolytic enzyme activity. *Marine Drugs* 12: 5277. DOI: 10.3390/md12105277.
- Kim H. 2017. Traditional Korean medicine made modern. In *Korea Magazine December 2017*. (Ed. by R. Koehler, G. Eaves & A. Bloom). Korean Culture and Information Service, Seoul. https://issuu.com/kocis9/docs/korea_magazine_1712; searched on 05 March 2019.
- Kim M.S., Kim J.Y., Choi W.H. & Lee S.S. 2008. Effects of seaweed supplementation on blood glucose concentration, lipid profile, and antioxidant enzyme activities in patients with type 2 diabetes mellitus. *Nutrition Research and Practice* 2: 62–67. DOI: 10.4162/nrp.2008.2.2.62.
- Ko S.-C., Kang M.C., Kang N., Kim H.-S., Lee S.-H., Ahn G., Jung W.-K. & Jeon Y.-J. 2017. Effect of angiotensin I-converting enzyme (ACE) inhibition and nitric oxide (NO) production of 6,6'-bieckol, a marine algal polyphenol and its anti-hypertensive effect in spontaneously hypertensive rats. *Process Biochemistry* 58: 326–332. DOI: 10.1016/j.procbio.2017.04.014.
- Koenig G. & Seneff S. 2015. Gamma-glutamyltransferase: A predictive biomarker of cellular antioxidant inadequacy and disease risk. *Disease Markers* Article ID 818570: 18 pp. DOI: 10.1155/2015/818570.
- Kraan S. 2012. Algal polysaccharides, novel applications and outlook. In: *Carbohydrates-comprehensive studies on glycobiology and glycotecnology* (Ed. by C.-F. Chang), pp. 489–532. Institute for New Technologies, Maastricht, Netherlands.

- Kumoro A., Johnny D. & Alfilovita D. 2016. Incorporation of microalgae and seaweed in instant fried wheat noodles manufacturing: nutrition and culinary properties study. *International Food Research Journal* 23: 715–722. [http://ifrj.upm.edu.my/23%20\(02\)%202016/\(36\).pdf](http://ifrj.upm.edu.my/23%20(02)%202016/(36).pdf); searched on 11 February 2019.
- Kurotani K., Akter S., Kashino I., Goto A., Mizoue T., Noda M., Sasazuki S., Sawada N. & Tsugane S. 2016. Quality of diet and mortality among Japanese men and women: Japan Public Health Center based prospective study. *British Medical Journal* 352: i1209. DOI: 10.1136/bmj.i1209.
- Lee S.H. & Jeon Y.J. 2015. Efficacy and safety of a dieckol-rich extract (AG-dieckol) of brown algae, *Ecklonia cava*, in pre-diabetic individuals: a double-blind, randomized, placebo-controlled clinical trial. *Food and Function* 6: 853–858. DOI: 10.1039/c4fo00940a.
- Lee Y.H., Yoon S.-J., Kim A., Seo H. & Ko S. 2016. Health performance and challenges in Korea: A review of the Global Burden of Disease Study 2013. *Journal of Korean Medical Science* 31: S114–S120. DOI: 10.3346/jkms.2016.31.S2.S114.
- Liu L., Heinrich M., Myers S. & Dworjanyn S.A. 2012. Towards a better understanding of medicinal uses of the brown seaweed *Sargassum* in traditional Chinese medicine: A phytochemical and pharmacological review. *Journal of Ethnopharmacology* 142: 591–619. DOI: 10.1016/j.jep.2012.05.046.
- LOGAN, A.C. (2003). Neurobehavioral aspects of omega-3 fatty acids: possible mechanisms and therapeutic value in major depression. *Alternative Medicine Review* 8: 410–425. <https://www.ncbi.nlm.nih.gov/pubmed>.

- Lokhande G., Carrow J.K., Thakur T., Xavier J.R., Parani M., Bayless K.J. & Gaharwar A.K. 2018. Nanoengineered injectable hydrogels for wound healing application. *Acta Biomaterialia* 70: 35–47. DOI: 10.1016/j.actbio.2018.01.045.
- López-López I., Bastida S., Ruiz-Capillas C., Bravo L., Larrea M., Sánchez-Muniz F., Cofrades S. & Jiménez-Colmenero F. 2009. Composition and antioxidant capacity of low-salt meat emulsion model systems containing edible seaweeds. *Meat Science* 83: 492–498. DOI: 10.1016/j.meatsci.2009.06.031.
- MacArtain P., Gill C.I.R., Brooks M., Campbell R. & Rowland I.R. 2007. Nutritional value of edible seaweeds. *Nutrition Reviews* 65: 535–543. DOI: 10.1111/j.1753-4887.2007.tb00278.x.
- Maruyama K., Iso H., Date C., Kikuchi S., Watanabe Y., Wada Y., Inaba Y. & Tamakoshi A. 2013. Dietary patterns and risk of cardiovascular deaths among middle-aged Japanese: JACC Study. *Nutrition, Metabolism and Cardiovascular Diseases* 23: 519–527. DOI: 10.1016/j.numecd.2011.10.007.
- Medina-Remón A., Kirwan R., Lamuela-Raventós R.M. & Estruch R. 2018. Dietary patterns and the risk of obesity, type 2 diabetes mellitus, cardiovascular diseases, asthma, and neurodegenerative diseases. *Critical Reviews in Food Science and Nutrition* 58: 262–296. DOI: 10.1080/10408398.2016.1158690.
- Misurcova L. 2011. Chemical composition of seaweeds. In: *Handbook of marine macroalgae: biotechnology and applied phycology* (Ed. by S.-K. Kim), pp. 171–192. John Wiley & Sons, Hoboken, New Jersey, USA.

- Moore L.E. 2018. Oral hypoglycemic agents. In *Diabetes in pregnancy: the complete guide to management*. (Ed. by L.E. Moore), pp. 103–110. Springer International Publishing, Cham, Switzerland.
- Murata M. & Nakazoe J. 2001. Production and use of marine algae in Japan. *Japan Agricultural Research Quarterly: JARQ* 35: 281–290. DOI: 10.6090/jarq.35.281.
- Murray M., Dordevic A., Ryan L. & Bonham M. 2018. The impact of a single dose of a polyphenol-rich seaweed extract on postprandial glycaemic control in healthy adults: a randomised cross-over trial. *Nutrients* 10: 270. DOI: 10.3390/nu10030270.
- Mysliwa-Kurdziel B. & Solymosi K. 2017. Phycobilins and phycobiliproteins used in food industry and medicine. *Mini-Reviews in Medicinal Chemistry* 17: 1173–1193. DOI: 10.2174/1389557516666160912180155.
- Namvar F., Mohamed S., Fard S.G., Behravan J., Mustapha N.M., Alitheen N.B.M. & Othman F. 2012. Polyphenol-rich seaweed (*Eucheuma cottonii*) extract suppresses breast tumour via hormone modulation and apoptosis induction. *Food Chemistry* 130: 376–382. DOI: 10.1016/j.foodchem.2011.07.054.
- Nanri A., Mizoue T., Shimazu T., Ishihara J., Takachi R., Noda M., Iso H., Sasazuki S., Sawada N., Tsugane S. & for the Japan Public Health Center-Based Prospective Study G. 2017. Dietary patterns and all-cause, cancer, and cardiovascular disease mortality in Japanese men and women: the Japan public health center-based prospective study. *PLOS ONE* 12: e0174848. DOI: 10.1371/journal.pone.0174848.
- Narasimhan M.K., Pavithra S.K., Krishnan V. & Chandrasekaran M. 2013. In vitro analysis of antioxidant, antimicrobial and antiproliferative activity of *Enteromorpha antenna*, *Enteromorpha linza* and *Gracilaria corticata* extracts. *Jundishapur Journal of Natural*

Pharmaceutical Products 8: 151–159.

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3941891/>.

NCD-RisC. 2016. NCD Risk Factor Collaboration. Worldwide trends in diabetes since 1980: a pooled analysis of 751 population-based studies with 4.4 million participants. *The Lancet* 387: 1513–1530. DOI: 10.1016/S0140-6736(16)00618-8.

Nelson S.M., Gao Y.-T., Nogueira L.M., Shen M.-C., Wang B., Rashid A., Hsing A.W. & Koshiol J. 2017. Diet and biliary tract cancer risk in Shanghai, China. *PLOS ONE* 12: e0173935. DOI: 10.1371/journal.pone.0173935.

OECD. 2017. Organisation for Economic Co-operation and Development. Obesity Update 2017. <https://www.oecd.org/els/health-systems/Obesity-Update-2017.pdf>; searched on 24 January 2019.

Olivero-David R., Schultz-Moreira A., Vázquez-Velasco M., González-Torres L., Bastida S., Benedí J., Isabel Sanchez-Reus M., José González-Muñoz M. & Sánchez-Muniz F.J. 2011. Effects of nori- and wakame-enriched meats with or without supplementary cholesterol on arylesterase activity, lipaemia and lipoproteinaemia in growing Wistar rats. *British Journal of Nutrition* 106: 1476–1486. DOI: 10.1017/S000711451100198X.

Paniagua-Michel J.d.J., Olmos-Soto J. & Morales-Guerrero E.R. 2014. Algal and microbial exopolysaccharides: new insights as biosurfactants and bioemulsifiers. In: *Advances in food and nutrition research* (Ed by S.-K. Kim), pp. 221–257. Academic Press, San Diego, California, USA.

Paradis M.-E., Couture P. & Lamarche B. 2011. A randomised crossover placebo-controlled trial investigating the effect of brown seaweed (*Ascophyllum nodosum* and *Fucus vesiculosus*)

- on postchallenge plasma glucose and insulin levels in men and women. *Applied Physiology, Nutrition, and Metabolism* 36: 913–919. DOI: 10.1139/h11-115.
- Park Y., Lee J., Oh J.H., Shin A. & Kim J. 2016. Dietary patterns and colorectal cancer risk in a Korean population: a case-control study. *Medicine* 95: e3759. DOI: 10.1097/MD.00000000000003759.
- Patterson E., Wall R., Fitzgerald G., Ross R. & Stanton C. 2012. Health implications of high dietary omega-6 polyunsaturated fatty acids. *Journal of Nutrition and Metabolism* 2012: 16. DOI: 10.1155/2012/539426.
- Pérez M.J., Falqué E. & Domínguez H. 2016. Antimicrobial action of compounds from marine seaweed. *Marine Drugs* 14: 52. DOI: 10.3390/md14030052.
- Pinteus S., Silva J., Alves C., Horta A., Fino N., Rodrigues A.I., Mendes S. & Pedrosa R. 2017a. Cytoprotective effect of seaweeds with high antioxidant activity from the Peniche coast (Portugal). *Food Chemistry* 218: 591–599. DOI: 10.1016/j.foodchem.2016.09.067.
- Pinteus S., Silva J., Alves C., Horta A., Thomas O. & Pedrosa R. 2017b. Antioxidant and cytoprotective activities of *Fucus spiralis* seaweed on a human cell *in vitro* model. *International Journal of Molecular Sciences* 18: 292. DOI: 10.3390/ijms18020292.
- Pongpichaiudom A. & Songsermpong S. 2018. Evaluation of microstructure and quality characteristics of microwave-dried instant noodles enriched with chicken meat, egg yolk, and seaweed. *Journal of Food Measurement and Characterization* 12: 22–34. DOI: 10.1007/s11694-017-9613-3.
- Pritchard M.F., Powell L.C., Jack A.A., Powell K., Beck K., Florance H., Forton J., Rye P.D., Dessen A., Hill K.E. & Thomas D.W. 2017. A low-molecular-weight alginate oligosaccharide disrupts pseudomonal microcolony formation and enhances antibiotic

- effectiveness. *Antimicrobial Agents and Chemotherapy* 61: e00762-17. DOI: 10.1128/AAC.00762-17.
- Qin Y. 2018. Applications of bioactive seaweed substances in functional food products. In: *Bioactive seaweeds for food applications: natural ingredients for healthy diets* (Ed by Y. Qin), pp. 111–134. Academic Press, San Diego, California, USA.
- Ramesh M. & Muthuraman A. 2018. Chapter 1 – Flavoring and coloring agents: health risks and potential problems. In: *Natural and artificial flavoring agents and food dyes* (Ed. by A.M. Grumezescu & A.M. Holban), pp. 1–28. Academic Press, San Diego, California, USA.
- Ren R., Gong J., Zhao Y., Zhuang X., Ye Y. & Lin W. 2017. Sulfated polysaccharides from *Enteromorpha prolifera* suppress SREBP-2 and HMG-CoA reductase expression and attenuate non-alcoholic fatty liver disease induced by a high-fat diet. *Food & Function* 8: 1899–1904.
- Ren R., Gong J., Zhao Y., Zhuang X., Ye Y., Huang F. & Lin W. 2018. Sulfated polysaccharide from *i* suppresses SREBP-1c and ACC expression to lower serum triglycerides in high-fat diet-induced hyperlipidaemic rats. *Journal of Functional Foods* 40: 722–728. DOI: 10.1016/j.jff.2017.12.010.
- Rhein-Knudsen N., Ale M.T. & Meyer A.S. 2015. Seaweed hydrocolloid production: an update on enzyme assisted extraction and modification technologies. *Marine Drugs* 13: 3340–3359. DOI: 10.3390/md13063340.
- Richter M.F. & Hergenrother P.J. 2018. The challenge of converting Gram-positive-only compounds into broad-spectrum antibiotics. *Annals of the New York Academy of Sciences* 1435: 18–38. DOI: 10.1111/nyas.13598.

- Rindi F., Soler-Vila A. & Guiry M.D. 2012. Taxonomy of marine macroalgae used as sources of bioactive compounds. In: *Marine bioactive compounds* (Ed. by M. Hayes), pp. 1–53. Springer, New York, New York, USA.
- Rodrigues D., Freitas A.C., Pereira L., Rocha-Santos T.A., Vasconcelos M.W., Roriz M., Rodríguez-Alcalá L.M., Gomes A.M. & Duarte A.C. 2015. Chemical composition of red, brown and green macroalgae from Buarcos Bay in central west coast of Portugal. *Food Chemistry* 183: 197–207. DOI: 10.1016/j.foodchem.2015.03.057.
- Roohinejad S., Koubaa M., Barba F.J., Saljoughian S., Amid M. & Greiner R. 2017. Application of seaweeds to develop new food products with enhanced shelf-life, quality and health-related beneficial properties. *Food Research International* 99: 1066–1083. DOI: 10.1016/j.foodres.2016.08.016.
- Roth G.A., Huffman M.D., Moran A.E., Feigin V., Mensah G.A., Naghavi M. & Murray C.J. 2015. Global and regional patterns in cardiovascular mortality from 1990 to 2013. *Circulation* 132: 1667–1678.
- Roy M.-C., Anguenot R., Fillion C., Beaulieu M., Bérubé J. & Richard D. 2011. Effect of a commercially-available algal phlorotannins extract on digestive enzymes and carbohydrate absorption *in vivo*. *Food Research International* 44: 3026–3029. DOI: 10.1016/j.foodres.2011.07.023.
- Rupérez P. 2002. Mineral content of edible marine seaweeds. *Food Chemistry* 79: 23–26. DOI: 10.1016/S0308-8146(02)00171-1.
- Sathya R., Kanaga N., Sankar P. & Jeeva S. 2017. Antioxidant properties of phlorotannins from brown seaweed *Cystoseira trinodis* (Forsskål) C. Agardh. *Arabian Journal of Chemistry* 10: S2608–S2614. DOI: 10.1016/j.arabjc.2013.09.039.

- Sato M., Oba T., Yamaguchi T., Nakano T., Kahara T., Funayama K., Kobayashi A. & Nakano T. 2002. Antihypertensive effects of hydrolysates of Wakame (*Undaria pinnatifida*) and their angiotensin-i-converting enzyme inhibitory activity. *Annals of Nutrition and Metabolism* 46: 259–267. DOI: 10.1159/000066495.
- Sellimi S., Ksouda G., Nasri R., Rinaudo M., Nasri M. & Hajji M. 2017. Enhancing colour and oxidative stabilities of reduced-nitrite turkey meat sausages during refrigerated storage using fucoxanthin purified from the Tunisian seaweed *Cystoseira barbata*. *Food and Chemical Toxicology* 107: 620–629. DOI: 10.1016/j.fct.2017.04.001.
- Shannon E. & Abu-Ghannam N. 2016. Antibacterial derivatives of marine algae: an overview of pharmacological mechanisms and applications. *Marine Drugs* 14: 81. DOI: 10.3390/md14040081.
- Shannon E. & Abu-Ghannam N. 2017. Optimisation of fucoxanthin extraction from Irish seaweeds by response surface methodology. *Journal of Applied Phycology* 29: 1027–1036. DOI: 10.1007/s10811-016-0983-4.
- Shannon E., Jaiswal A. & Abu-Ghannam N. 2018. Polyphenolic content and antioxidant capacity of white, green, black, and herbal teas: a kinetic study. *Food Research* 2: 1–11. DOI: 10.26656/fr.2017.2(1).117.
- Sharifuddin Y., Chin Y.-X., Lim P.-E. & Phang S.-M. 2015. Potential bioactive compounds from seaweed for diabetes management. *Marine Drugs* 13: 5447–5491. DOI: 10.3390/md13085447.
- Shimizu H., Ross R.K., Bernstein L., Yatani R., Henderson B.E. & Mack T.M. 1991. Cancers of the prostate and breast among Japanese and white immigrants in Los Angeles County. *British Journal Of Cancer* 63: 963. DOI: 10.1038/bjc.1991.210.

- Shin H.C., Kim S.H., Park Y., Lee B.H. & Hwang H.J. 2012. Effects of 12-week oral supplementation of *Ecklonia cava* polyphenols on anthropometric and blood lipid parameters in overweight Korean individuals: a double-blind randomized clinical trial. *Phytotherapy Research* 26: 363–368. DOI: 10.1002/ptr.3559.
- Simopoulos A.P. 2016. An increase in the omega-6/omega-3 fatty acid ratio increases the risk for obesity. *Nutrients* 8: 128. DOI: 10.3390/nu8030128.
- Song W., Wang Z., Zhang X. & Li Y. 2018. Ethanol extract from *Ulva prolifera* prevents high-fat diet-induced insulin resistance, oxidative stress, and inflammation response in mice. *BioMed Research International*: Article ID 1374565. 9. DOI: 10.1155/2018/1374565.
- Stephen A.M., Champ M.M.J., Cloran S.J., Fleith M., van Lieshout L., Mejbourn H. & Burley V.J. 2017. Dietary fibre in Europe: current state of knowledge on definitions, sources, recommendations, intakes and relationships to health. *Nutrition Research Reviews* 30: 149–190. DOI: 10.1017/S095442241700004X.
- Suetsuna K. & Nakano T. 2000. Identification of an antihypertensive peptide from peptic digest of wakame (*Undaria pinnatifida*). *The Journal of Nutritional Biochemistry* 11: 450–454. DOI: 10.1016/S0955-2863(00)00110-8.
- Tanemura Y., Yamanaka-Okumura H., Sakuma M., Nii Y., Taketani Y. & Takeda E. 2014. Effects of the intake of *Undaria pinnatifida* (Wakame) and its sporophylls (Mekabu) on postprandial glucose and insulin metabolism. *The Journal of Medical Investigation* 61: 291–297. DOI: 10.2152/jmi.61.291.
- Teas J., Baldeón M.E., Chiriboga D.E., Davis J.R., Sarriés A.J. & Braverman L.E. 2009. Could dietary seaweed reverse the metabolic syndrome? *Asia Pacific Journal of Clinical Nutrition* 18: 145–154. <https://www.ncbi.nlm.nih.gov/pubmed/19713172>.

- Thompson P.D., Panza G., Zaleski A. & Taylor B. 2016. Statin-associated side effects. *Journal of the American College of Cardiology* 67: 2395–2410. DOI: 10.1016/j.jacc.2016.02.071.
- Turck D., Bresson J.L., Burlingame B., Dean T., Fairweather-Tait S., Heinonen M., Hirsch-Ernst K.I., Mangelsdorf I., McArdle H., Naska A., Neuhäuser-Berthold M., Nowicka G., Pentieva K., Sanz Y., Siani A., Sjödin A., Stern M., Tomé D., Vinceti M., Willatts P., Engel K.H., Marchelli R., Pöting A., Poulsen M., Salminen S., Schlatter J., Arcella D., Gelbmann W., Sesmaisons-Lecarré A., Verhagen H. & Loveren H. 2016. Guidance on the preparation and presentation of an application for authorisation of a novel food in the context of Regulation (EU) 2015/2283. Section 2.2.3. Foods consisting of, isolated from or produced from microorganisms, fungi or algae. *EFSA Journal* 14: e04594. DOI: 10.2903/j.efsa.2016.4594.
- U.S.F.D.A. 2017. Code of Federal Regulations Title 21. Subchapter B - food for human consumption. PART 184 Direct food substances affirmed as generally recognized as safe. Subpart B listing of specific substances affirmed as GRAS. Sec. 184.1120 Brown algae. <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?fr=184.1120>; searched on 15 February 2019.
- Uchida M., Kurushima H., Hideshima N., Araki T., Ishihara K., Murata Y., Touhata K. & Ishida N. 2018. Preparation and characterization of fermented seaweed sauce manufactured from low-quality nori (dried and fresh fronds of *Pyropia yezoensis*). *Fisheries Science* 84: 589–596. DOI: 10.1007/s12562-018-1184-7.
- Valko M., Jomova K., Rhodes C.J., Kuča K. & Musílek K. 2016. Redox-and non-redox-metal-induced formation of free radicals and their role in human disease. *Archives of Toxicology* 90: 1–37. DOI: 10.1007/s00204-015-1579-5.

- Vogel V.G. 2018. Epidemiology of Breast Cancer. In: *The breast* (Ed. by K.I. Bland, E.M. Copeland, V.S. Klimberg & W.J. Gradishar), pp. 207–218.e4. Elsevier, Philadelphia, Pennsylvania, USA.
- W.H.O. 2019. World Health Organization. Double burden of malnutrition. <https://www.who.int/nutrition/double-burden-malnutrition/en/>; searched on 13 April 2019.
- W.H.O. 2016a. World Health Organization. Diabetes country profiles 2016 - United Kingdom. http://www.who.int/diabetes/country-profiles/gbr_en.pdf?ua=1; searched on 28 January 2019.
- W.H.O. 2016b. World Health Organization. Diabetes country profiles 2016 - United States of America. http://www.who.int/diabetes/country-profiles/usa_en.pdf?ua=1; searched on 16 January 2019.
- W.H.O. 2017. Global Health Observatory (GHO) data. Raised blood pressure. Situation and trends. http://www.who.int/gho/ncd/risk_factors/blood_pressure_prevalence_text/en/; searched on 20 January 2019.
- Wei Y., Liu Q., Xu C., Yu J., Zhao L. & Guo Q. 2016. Damage to the membrane permeability and cell death of *Vibrio parahaemolyticus* caused by phlorotannins with low molecular weight from *Sargassum thunbergii*. *Journal of Aquatic Food Product Technology* 23: 323–333. DOI: 10.1080/10498850.2013.851757.
- Willyard C. 2017. The drug-resistant bacteria that pose the greatest health threats. *Nature* 543: 15. DOI: 10.1038/nature.2017.21550.
- Wong K. & Cheung P.C. 2000. Nutritional evaluation of some subtropical red and green seaweeds: Part I—proximate composition, amino acid profiles and some physico-chemical properties. *Food Chemistry* 71: 475–482. DOI: 10.1016/S0308-8146(00)00176-X.

- Wong K.H. & Cheung P.C.K. 2001. Nutritional evaluation of some subtropical red and green seaweeds Part II. *In vitro* protein digestibility and amino acid profiles of protein concentrates. *Food Chemistry* 72: 11–17. DOI: 10.1016/S0308-8146(00)00176-X.
- Yamazaki D., Hitomi H. & Nishiyama A. 2018. Hypertension with diabetes mellitus complications. *Hypertension Research* 41: 147-156. DOI: 10.1038/s41440-017-0008-y.
- Yamori Y., Sagara M., Arai Y., Kobayashi H., Kishimoto K., Matsuno I., Mori H. & Mori M. 2017. Soy and fish as features of the Japanese diet and cardiovascular disease risks. *PLoS ONE* 12: e0176039. DOI: 10.1371/journal.pone.0176039.
- Yan X., Chuda Y., Suzuki M. & Nagata T. 1999. Fucoxanthin as the major antioxidant in *Hijikia fusiformis*, a common edible seaweed. *Bioscience, Biotechnology, and Biochemistry* 63: 605–607. DOI: 10.1271/bbb.63.605.
- Yang Y.J., Nam S.-J., Kong G. & Kim M.K. 2010. A case–control study on seaweed consumption and the risk of breast cancer. *British Journal of Nutrition* 103: 1345–1353. DOI: 10.1017/S0007114509993242.
- Zaragoza M.C., López D., P. Sáiz M., Poquet M., Pérez J., Puig-Parellada P., Marmol F., Simonetti P., Gardana C. & Lerat Y. 2008. Toxicity and antioxidant activity *in vitro* and *in vivo* of two *Fucus vesiculosus* extracts. *Journal of Agricultural and Food Chemistry* 56: 7773–7780. DOI: 10.1021/jf8007053.
- Zerrifi S., El Khalloufi F., Oudra B. & Vasconcelos V. 2018. Seaweed bioactive compounds against pathogens and microalgae: Potential uses on pharmacology and harmful algae bloom control. *Marine Drugs* 16: 55. DOI:10.3390/md16020055.
- Zhao C., Yang C., Chen M., Lv X., Liu B., Yi L., Cornara L., Wei M.C., Yang Y.C., Tundis R. & Xiao J. 2018. Regulatory efficacy of brown seaweed *Lessonia nigrescens* extract on the

- gene expression profile and intestinal microflora in type 2 diabetic mice. *Molecular Nutrition & Food Research* 62: 1700730. DOI: 10.1002/mnfr.201700730.
- Zhou C., Elshkaki A. & Graedel T.E. 2018. Global human appropriation of net primary production and associated resource decoupling: 2010-2050. *Environmental Science & Technology* 52: 1208–1215. DOI: 10.1021/acs.est.7b04665.
- Ziegler R.G., Hoover R.N., Pike M.C., Hildesheim A., Nomura A.M., West D.W., Wu-Williams A.H., Kolonel L.N., Horn-Ross P.L. & Rosenthal J.F. 1993. Migration patterns and breast cancer risk in Asian-American women. *JNCI: Journal of the National Cancer Institute* 85: 1819–1827. DOI: 10.1093/jnci/85.22.1819.
- Zollman M., Robin A., Prabhu M., Polikovsky M., Gillis A, Greiserman S. & Goldberg A. Green technology in green macroalgal biorefineries. *Phycologia* 58: 516–534. DOI: 10.1080/00318884.2019.1640516.