Synthesis and Antimicrobial Evaluation of Carbohydrate and Polyhydroxylated Non-carbohydrate Fatty Acid Ester and Ether Derivatives

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Synthesis and antimicrobial evaluation of carbohydrate and polyhydroxylated non-carbohydrate fatty acid ester and ether derivatives.

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Abstract

A series of fatty acid ester and ether derivatives have been chemically synthesised based on carbohydrate and non-carbohydrate polyhydroxylated scaffolds. The synthesised compounds, along with their corresponding fatty acid monoglyceride antimicrobials, were evaluated for antimicrobial activity against Staphylococcus aureus and Escherichia coli. Of the derivatives synthesised several of the carbohydrate based compounds have antimicrobial efficacy comparable with commercially available antimicrobials. The results suggest that the nature of the carbohydrate core plays a role in the efficacy of carbohydrate fatty acid derivatives as antimicrobials.

Keywords

Fatty acid derivatives, lauric acid, monolaurin, antimicrobial activity, Staphylococcus aureus and Escherichia coli.
1. Introduction

The antimicrobial effects of fatty acids have been well documented.\(^1\) Generally, long chain fatty acids have activity against Gram-positive bacteria while short chain fatty acids are more active against Gram-negative bacteria. Lauric acid (medium chain fatty acid) is regarded as the most active, with reported activity against both Gram-positive and Gram-negative bacteria.\(^2\) Lauric acid and gentamicin combined have been reported to show activity against MRSA.\(^3\) Lauric acid is inexpensive and therefore may be very useful for infection control in hospitals.

Esterification of fatty acids with monohydric alcohols such as methanol or ethanol has been shown to reduce their antimicrobial activity.\(^4\) In contrast, esterification of fatty acids to the polyhydric alcohol glycerol increased their effectiveness.\(^5\) One of the most active of these antimicrobial derivatives is monolaurin (Lauricidin\(^\text{@}\)), the glycerol monoester of lauric acid, which is used as a key ingredient of antimicrobial food additives to inhibit the growth of undesirable microorganisms.\(^6,7\)

More recently, a study has shown that the corresponding ether of monolaurin, dodecylglycerol, had greater potency against *Streptococcus faecium* than monolaurin itself, albeit depending on the incubation conditions.\(^8\) The greater potency of dodecylglycerol was ascribed to its greater retention by the cell, and its action on specific receptors or enzymes.

Another class of fatty acid derivatives which have broad applications in the food industry are carbohydrate fatty acid esters.\(^9,10\) While they are most commonly employed as surfactants, their antimicrobial properties have been documented.\(^11\) The use of carbohydrate esters is increasingly favoured since they are biodegradable, are not harmful to the environment and they are non-toxic.\(^12\)
The most common carbohydrate fatty acid ester utilised to date is sucrose ester. They are commercially available and used for a variety of food applications. Kato and Shibasaki (1975) showed that the sucrose ester of lauric acid had potent antimicrobial activity against certain Gram-positive bacteria and fungi. They further showed that, in contrast to findings with glycerides, the diester of sucrose was more active, than the monoester. Of the diesters tested, sucrose dicaprylate showed the highest activity. Other oligosaccharide fatty acid esters, including maltose and maltotriose, have been synthesised. These sugar esters were shown to inhibit the growth of *Streptococcus sobrinus*, and are therefore potentially of significant value in the development of oral-hygiene products. One study investigating the effect of carbohydrate monoesters reported that among those synthesised, galactose laurate, fructose laurate and the reducing 6-**O-lauroylmannose** showed the highest inhibitory effect against *Streptococcus mutans*, while other analogs of hexose laurates showed no activity. This finding strongly suggests that the carbohydrate moiety can markedly affect the antimicrobial activity of the fatty acid and therefore further investigation is merited.

Recent work in the area of carbohydrate fatty acid esters has focused on establishing an effective regioselective, enzyme catalysed, synthesis of sugar derivatives for use as surfactants for industrial applications, however relatively few studies have examined role of the carbohydrate in antimicrobial activity. This study is concerned with the synthesis of carbohydrate and polyhydroxylated non-carbohydrate fatty acid derivatives for evaluation as antibacterial agents, with a view to examining the effect of variation of the hydrophilic moiety on antimicrobial activity. Therefore, we designed chemical syntheses to investigate the effects of carbohydrate versus non-carbohydrate hydrophilic cores, the number of fatty acids attached to the hydrophilic core, the monosaccharide core itself (and the anomeric
configuration with respect to glucopyranoside), the glycoconjugate linkage and the length of fatty acid chain on antimicrobial activity.

A quantitative assay for antimicrobial activity was used to allow comparisons between compounds and all were measured relative to the free fatty acids and monolaurin as reference compounds.

Enzymatic synthesis of novel sugar fatty acid esters has been widely employed and can be highly regioselective, although for some carbohydrates minor regiomeric isomers may be obtained. For this study, we have developed a chemical route to allow us synthesise a number of pure, regio-defined, monosaccharide mono fatty acid esters (Scheme 1). We have also developed a route to the corresponding ether derivatives (Scheme 2). In order to establish whether a second fatty acid conjugated to a monosaccharide would improve antimicrobial activity, a route was developed to synthesise a di-laurate derivative (Scheme 3). Furthermore, to investigate whether the structure and therefore the synthesis, could be simplified and retain activity, non-carbohydrate hydroxylated esters based on a pentaerythritol core were synthesised by a straightforward esterification (Scheme 4).

2. Results and Discussion

2.1 Synthesis

A designed chemical route to obtain mono-ester sugars is shown in Scheme 1 and is based on the following carbohydrate starting materials: 1a methyl α-D-glucopyranoside, 1b methyl β-D-glucopyranoside, 1c methyl α-D-mannopyranoside and 1d methyl α-D-galactopyranoside. The synthesis commenced with the selective protection of the primary hydroxyl of sugars 1a-d with a triisopropylsilyl (TIPS) group. The silyl derivatives were then fully protected with benzyl groups to give 2a-d. The removal of the TIPS group by tetrabutylammonium
fluoride in THF allowed for the esterification of the free 6-OH position with either lauroyl chloride to yield 3a-d or octanoyl chloride to yield 5a. Removal of the benzyl groups by catalytic hydrogenation led to the unprotected carbohydrate esters 4a-d and 6a respectively.

Table 1 Percentage yields of compounds 2a-d, 3a-d, 4a-d, 5a and 6a.

<table>
<thead>
<tr>
<th>Carbohydrate</th>
<th>2,3,4-tri-O-Bn-6-O-TIPS (1)</th>
<th>2,3,4-tri-O-Bn-6-O-lauroyl (2)</th>
<th>6-O-lauroyl (3)</th>
<th>2,3,4-tri-O-Bn-6-O-octanoyl (4)</th>
<th>6-O-octanoyl (5)</th>
<th>6a</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1a)</td>
<td>2a</td>
<td>3a</td>
<td>4a</td>
<td>5a</td>
<td>6a</td>
<td></td>
</tr>
<tr>
<td></td>
<td>85%</td>
<td>72%</td>
<td>86%</td>
<td>63%</td>
<td>73%</td>
<td></td>
</tr>
<tr>
<td>(1b)</td>
<td>2b</td>
<td>3b</td>
<td>4b</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>80%</td>
<td>70%</td>
<td>75%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(1c)</td>
<td>2c</td>
<td>3c</td>
<td>4c</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>51%</td>
<td>64%</td>
<td>75%</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(1d)</td>
<td>2d</td>
<td>3d</td>
<td>4d</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>50%</td>
<td>60%</td>
<td>86%</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Synthesis of the ether derivatives also commenced with the protection of the primary hydroxyl with a triisopropylsilyl group (Scheme 2). The sugars were then fully protected using paramethoxybenzyl chloride (PMB), to yield 7a-b. Removal of the TIPS group gave the free primary hydroxyl. Next, the lauric ether group was attached using dodecanyl chloride to give the fully protected ether derivatives 8a-b. Finally
oxidative cleavage of the PMB groups with CAN gave the mono-dodecanyl sugars 9a-b.


**Table 2** Percentage yields of compounds 7a-b, 8a-b and 9a-b.

<table>
<thead>
<tr>
<th>Carbohydrate</th>
<th>2,3,4-tri-O-PMB-6-O-TIPS (7)</th>
<th>2,3,4-tri-O-PMB-6-O-dodecanyl (8)</th>
<th>6-O-dodecanyl (9)</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(1a)</td>
<td>7a 59%</td>
<td>8a 50%</td>
<td>9a 73%</td>
</tr>
<tr>
<td>(1b)</td>
<td>7b 61%</td>
<td>8b 85%</td>
<td>9b 76%</td>
</tr>
</tbody>
</table>

The method used to synthesise di-lauroyl derivative 12a is shown in Scheme 3. The 4 and 6-OH positions of methyl α-D-glucopyranoside 1a were protected with a benzyldiene group using benzaldehyde dimethylacetal. The remaining free OH’s
were then converted to benzyl ethers to give 10a. Removal of the benzylidene acetal using catalytic TsOH in MeOH then enabled the esterification of the 4 and 6-OH to give 11a. Finally, removal of the benzyl groups by catalytic hydrogenation gave the diester derivative 12a.

**Scheme 3.** Reagents and Conditions: (i) pTSA, PhCH(OMe)₂, MeCN anhydr., rt. (ii) DMF anhydr., NaH, BnBr, rt. (95% yield over 2 steps) (iii) MeOH, TsOH. (iv) Pyr anhydr., DMAP, Lauroyl Cl, rt. (38% yield over 2 steps) (v) EtOH, Pd, H₂. (75% yield)

Direct esterification of pentaerythritol 13 using lauroyl chloride and DMAP in pyridine, yielded the non-sugar derivatives 14 and 15, shown in **Scheme 4**.

**Scheme 4.** Reagents and Conditions: (i) Pyr anhydr., DMAP, Lauroyl Cl, rt. (14 14%, 15 29%)

### 2.2 Antimicrobial activity of fatty acid derivatives

Two non-carbohydrate polyhydroxylated fatty acid ester derivatives, six carbohydrate fatty acid ester derivatives and two carbohydrate long chain alkyl ether derivatives,
together with their corresponding polyhydric alcohols, fatty acids and monoglycerides as controls, were tested against a Gram-positive bacteria, *Staphylococcus aureus*, and a Gram-negative bacteria, *Escherichia coli*, to assess their antimicrobial activity. The efficacy of the derivatives and controls were compared using Minimum Inhibitory Concentration values (MIC), which was defined as the lowest concentration of compound that showed no increase in cell growth for all the replicates compared to a negative control after 18 hours.

The polyhydric alcohols (carbohydrates and pentaerythritol) showed no antimicrobial activity or growth promoting effects for the microorganisms under the conditions used (results not shown).

### Table 3 MIC values of Fatty Acid Derivatives and Controls

<table>
<thead>
<tr>
<th>Compound</th>
<th><em>S. aureus</em> ATCC 25923</th>
<th><em>E. coli</em> ATCC 25922</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lauric acid</td>
<td>0.63 mM</td>
<td>10 mM</td>
</tr>
<tr>
<td>Monolaurin</td>
<td>0.04 mM</td>
<td>20 mM</td>
</tr>
<tr>
<td>Caprylic acid</td>
<td>5 mM</td>
<td>12.5 mM</td>
</tr>
<tr>
<td>Monocaprylin</td>
<td>2.5 mM</td>
<td>6.25 mM</td>
</tr>
<tr>
<td>Methyl 6-<em>O</em>-lauroyl-<em>α</em>-D-glucopyranoside (4a)</td>
<td>0.31 mM</td>
<td>20 mM</td>
</tr>
<tr>
<td>Methyl 6-<em>O</em>-lauroyl-<em>β</em>-D-glucopyranoside (4b)</td>
<td>0.04 mM</td>
<td>20 mM</td>
</tr>
<tr>
<td>Methyl 6-<em>O</em>-octanoyl-<em>α</em>-D-glucopyranoside (6a)</td>
<td>2.5 mM</td>
<td>12.5 mM</td>
</tr>
<tr>
<td>Methyl 6-<em>O</em>-dodecanyl-<em>α</em>-D-glucopyranoside (9a)</td>
<td>0.04 mM</td>
<td>20 mM</td>
</tr>
<tr>
<td>Methyl 6-<em>O</em>-dodecanyl-<em>β</em>-D-glucopyranoside (9b)</td>
<td>2.5 mM</td>
<td>20 mM</td>
</tr>
<tr>
<td>Methyl 4,6-di-<em>O</em>-lauroyl-<em>α</em>-D-glucopyranoside (12a)</td>
<td>ND*</td>
<td>ND</td>
</tr>
<tr>
<td>Methyl 6-<em>O</em>-lauroyl-<em>α</em>-D-mannopyranoside (4c)</td>
<td>0.04 mM</td>
<td>20 mM</td>
</tr>
<tr>
<td>Methyl 6-<em>O</em>-lauroyl-<em>α</em>-D-galactopyranoside (4d)</td>
<td>&gt;10 mM</td>
<td>&gt;20 mM</td>
</tr>
<tr>
<td>Mono lauroyl pentaerythritol (14)</td>
<td>&gt;10 mM</td>
<td>&gt;20 mM</td>
</tr>
<tr>
<td>Di lauroyl pentaerythritol (15)</td>
<td>ND</td>
<td>ND</td>
</tr>
</tbody>
</table>

* Not determined due to insolubility
The data in Table 3 show that the monoglycerides monolaurin and monocaprylin, had
greater activity compared to the free fatty acids lauric acid and caprylic acid against S.
aureus. Of the monoglycerides and free fatty acids tested, monolaurin had the lowest
MIC values for S. aureus, with a value of 0.04 mM compared to a value of 0.63 mM
for lauric acid. Furthermore, monocaprylin showed MIC values of 2.5 mM against S.
aureus compared to the value of 5.0 mM for caprylic acid. With respect to E. coli,
monolaurin showed less inhibitory effect than lauric acid with values of 20 mM and
10 mM respectively. In contrast, monocaprylin showed activity against E. coli at
concentrations of 6.25 mM compared with caprylic acid value of 12.5 mM.
All fatty acid derivatives showed greater antimicrobial activity against S. aureus than
E. coli.
Among the sugar fatty acid esters and the sugar alkyl ethers prepared, methyl 6-O-
dodecanyl-α-D-glucopyranoside 9a, methyl 6-O-lauroyl-α-D-mannopyranoside 4c and
methyl 6-O-lauroyl-β-D-glucopyranoside 4b showed the best inhibitory effects for S.
aureus, with MIC values of 0.04 mM. The next derivative in order of efficacy was
methyl 6-O-lauroyl-α-D-glucopyranoside 4a, with a value of 0.31 mM. Methyl 6-O-
octanoyl-α-D-glucopyranoside 6a was comparable to monocaprylin against S. aureus
with values of 2.5 mM. This compound was also more active than any of the lauric
acid derivatives against E. coli. Methyl 6-O-dodecanyl-β-D-glucopyranoside 9b gave
similar results to 6a for S. aureus with values of 2.5 mM. The galactopyranoside ester
derivative 4d and the mono-lauroyl pentaerythritol 14, were the least active
compounds tested, both with comparatively negligible MIC values of >10 mM for S.
aureus and >20mM for E. coli.
The di-substituted methyl 4,6-di-O-lauroyl-α-D-glucopyranoside 12a did not show
any activity comparable with either the monoglycerides or indeed the mono-
substituted sugar derivatives. This was attributed to poor solubility in water, as was the case for the di-substituted non-sugar compound di-lauroyl pentaerythritol 15.

2.3 Discussion

In this present study, we have evaluated the effect of polyhydroxylated fatty acid derivatives as inhibitors of a Gram-positive (S. aureus) and a Gram-negative (E. coli) microorganism of concern to the food and healthcare industries. Several of the synthesised compounds have antimicrobial efficacy comparable with commercially available antimicrobials against S. aureus. We studied the effect of carbohydrate versus non-carbohydrate hydrophilic cores (carbohydrate and pentaerythritol laurates), the degree of substitution (monoester and diester), the monosaccharide core (glucopyranoside, mannopyranoside and galactopyranoside), the anomic configuration (α and β glucopyranoside), the type of fatty acid carbohydrate linkage (ester and ether), and the length of fatty acid chain (lauric and caprylic) on antimicrobial activity.

As with the monoglycerides and free fatty acids, all of the fatty acid derivatives that were found to be active showed greater antimicrobial activity against the S. aureus than E. coli.

The non-carbohydrate pentaerythritol monoester 14, which has the same number of free hydroxyl groups as the carbohydrate monoester derivatives, showed negligible activity against both microorganisms tested, indicating that the carbohydrate itself could play an important role in the antimicrobial activity of these compounds.
The degree of substitution of these derivatives was also shown to be crucial as both the non-sugar pentaerythritol diester 15 and the carbohydrate methyl α-D-glucopyranoside diester 12a were much less soluble in water than the monoesters. As a consequence, no antimicrobial activity results for these compounds could be obtained.

With regard to the influence of different sugar cores, the results showed that the lauric ester derivative of methyl α-D-mannopyranoside 4c and methyl β-D-glucopyranoside 4b, showed higher activity than any other ester derivatives against S. aureus, supporting the observation that the nature of the carbohydrate is involved in the antimicrobial efficacy of the derivatives. This conclusion is consistent with results of an earlier study by Watanabe et al.15

Further evidence for this is noted in the results for the lauric ester anomers of methyl glucopyranoside 4a and 4b. A difference was noted when these compounds were tested against S. aureus with the beta configuration showing higher activity. The lauric ether anomers of methyl glucopyranoside 9a and 9b also showed a marked difference in activity when tested against S. aureus, with the alpha configuration showing a much higher activity.

In addition, the difference in activity between the ester and ether conjugates of the same carbohydrate showed that for the methyl α-D-glucopyranoside derivatives, the ether derivative 9a was more active than the ester 4a, however for methyl β-D-glucopyranoside, the ester 4b was more active than the ether 9b. These results indicate that, in combination with other factors, the nature of the bond conjugating the fatty acid to the carbohydrate could play some role in antimicrobial activity.

The importance of the chain length of the fatty acid ester was investigated using both lauric and caprylic derivatives. The lauric ester derivative 4a showed much higher
activity against *S. aureus* compared to the corresponding caprylic ester derivative 6a. Conversely, the caprylic ester derivative 6a showed higher activity against *E. coli*, compared with the lauric derivative 4a. This trend was also observed for the monoglyceride controls and is in accordance with general trends observed for medium and short chain fatty acids. 2

In conclusion, these results suggest that the nature of the carbohydrate core plays a role in the efficacy of carbohydrate fatty acid derivatives as antimicrobials, and therefore further optimisation may be possible. However, to confirm the trends outlined with respect to the importance of the carbohydrate moiety and the role of the nature of the glycoconjugate bond, further studies are warranted using a wider range of Gram-positive and Gram-negative microorganisms, which would allow for evaluation of potential species and strain effects.

3. Experimental

3.1 Synthesis

3.1.1 General methods

All air and moisture-sensitive reactions were performed under an inert nitrogen atmosphere. All reactions performed under a hydrogen atmosphere were performed in a Parr Hydrogenator Apparatus. Anhydrous DMF, THF, Pyridine and MeCN were purchased from Sigma Aldrich. TLC was performed on aluminium sheets precoated with Silica Gel 60 (HF<sub>254</sub>, Fluka) and spots visualised by UV and charring with H<sub>2</sub>SO<sub>4</sub>-EtOH (1:20). Flash Column Chromatography was carried out with Silica Gel 60 (0.040-0.630 mm, E. Merck) and using stepwise solvent polarity gradient correlated with TLC mobility. Chromatography solvents used were EtOAc (Riedel-deHaen), MeOH (Riedel-deHaen) and petroleum ether (b.p. 40-60 °C, Fluka). Optical rotations were determined with an AA-% Series Optical Activity Ltd Polarimeter.
NMR spectra were recorded with Varian Inova 300 and Varian NMRAS 400 spectrometers. Chemical shifts are reported relative to internal Me₄Si in CDCl₃ (δ 0.0) for ¹H and CDCl₃ (δ 77.0) for ¹³C. Coupling constants are reported in hertz.

FTIR spectra were recorded with a Nicolet FT-IR 5DXB infrared spectrometer, samples were prepared in a KBr matrix. Low resolution mass spectra were measured on a Quatromicro tandem quadropole mass spectrometer. Methyl-α-D-glucopyranoside, methyl-β-D-glucopyranoside, methyl-α-D-mannopyranoside, methyl-α-D-galactopyranoside, pentaerythritol, 1-chlorododecane, lauroyl chloride and octanoyl chloride were purchased from Sigma Aldrich.

3.1.2 Methyl 2,3,4-tri-O-benzyl-6-O-trisopropylsilyl-α-D-glucopyranoside (2a)

A solution of 1a (5 g, 25 mmol) in DMF anhydrous (120 mL) was treated with trisopropylsilyl chloride (15 mL, 75 mmol) and imidazole (5 g, 75 mmol) and allowed to stir at room temperature for 24 h. The crude TIPS protected intermediate was then concentrated in vacuo and dissolved in EtOAc. It was washed with 10% HCl, water, followed by sat. aq. NaHCO₃, and finally sat. aq. NaCl. It was then dried over anhydrous MgSO₄, and concentrated under reduced pressure. The crude product was dissolved in DMF anhydrous (50 mL) and cooled to 0 °C. NaH (5 g, 125 mmol) was added portion wise, BnBr (9 mL, 75 mmol) was added and the mixture was allowed to warm to room temperature and stir for 24 h. MeOH (50 mL) was added to quench the mixture which was stirred for 1 h. The fully protected sugar was then concentrated in vacuo and dissolved in EtOAc. The solution was washed with water, dried over anhydrous MgSO₄, and concentrated under diminished pressure. The resulting residue was purified by chromatography (petroleum ether-EtOAc) to give 2a (13.2 g, 85%); [α]D 10.7° (c 0.07, CHCl₃); FTIR (KBr): 2923, 1733, 1498, 1455, 909, 884, 791, 695 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.27 (ms, 15H,
aromatic H), 4.91, (AB d, 2H, J 11.0, OCH$_2$Ph), 4.78, (AB d, 2H, J 11.0, OCH$_2$Ph), 4.74 (AB d, 2H, J 12.0, OCH$_2$Ph), 4.61 (d, 1H, J$_{1,2}$ 3.5, H-1), 3.99 (apt t, 1H, J$_{3,4}$ 9.5, J$_{3,5}$ 9.5, H-3), 3.84 (d, 2H, J$_{5,6}$ 4.5, H-6a,6b), 3.64 (m, 1H, H-5), 1.10-1.02 (ms, 18H, each TIPS CH$_3$), 0.88 (m, 3H, each TIPS CH); 13C NMR (CDCl$_3$): δ 139.1, 138.7, 138.5 (each s, each aromatic C), 128.65, 128.63, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8 (each d, each aromatic CH), 98.0 (d, C-1), 82.5, 80.5, 78.1, 76.1 (each d), 76.1, 75.3, 73.6 (each t, each CH$_2$Ph), 62.9 (t, C-6), 55.0 (q, OCH$_3$), 18.3, 18.2 (each q, each TIPS CH$_3$), 12.2 (each d, each TIPS CH); LRMS: Found, 643.3; required, 643.9; [M + Na]$^+$.

3.1.3 Methyl 2,3,4-tri-O-benzyl-6-O-triisopropylsilyl-β-D-glucopyranoside (2b)

Treatment of 1b (4.5 g, 23.17 mmol) as described for 1a gave 2b (8.7 g, 80%); [α]$_D$ 23º (c 0.01, CHCl$_3$); FTIR (KBr): 2863, 1730, 1497, 1454, 1399, 1277, 882, 802, 751, 697. cm$^{-1}$; 1H NMR (400 MHz, CDCl$_3$): δ 7.37-7.28 (ms, 15H, aromatic H), 4.90, 4.88, 4.83 (each AB d, 6H, J 11.0, OCH$_2$Ph), 4.30 (d, 1H, J$_{1,2}$ 7.5, H-1), 4.00-3.90 (overlapping signals, 3H, H-5,6), 3.66 (m, 1H, H-3), 3.53 (s, 3H, OCH$_3$), 3.41 (m, 1H, H-2), 3.34 (m, 1H, H-4), 1.26-1.05 (ms, 21H, TIPS); 13C NMR (CDCl$_3$): δ 138.98, 138.92, 138.7 (each s, each aromatic C), 128.69, 128.65, 128.62, 128.5, 128.3, 128.2, 128.0, 127.9, 127.8 (each d, each aromatic CH), 104.7 (d, C-1), 84.9, 82.9, 77.8, 76.2 (each d), 76.0, 75.3, 75.0 (each t, each CH$_2$Ph), 62.7 (t, C-6), 56.9 (q, OCH$_3$), 18.3, 18.2 (each q, each TIPS CH$_3$), 12.3 (d, TIPS CH); LRMS: Found, 643.3 required, 643.9 [M + Na]$^+$.

3.1.4 Methyl 2,3,4-tri-O-benzyl-6-O-triisopropylsilyl-α-D-mannopyranoside (2c)

Treatment of 1c (4 g, 20 mmol) as described for 1a gave 2c (6.5 g, 51%); [α]$_D$ 25.5º (c 0.05, CHCl$_3$); FTIR (KBr): 3056, 2864, 1496, 1363, 1324, 970, 882, 790, 734, 696
1 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): ²δ 7.38-7.24 (multiple signals, 15H, each aromatic H), 4.79 (AB d, 2H, J 11.0, OCH₂Ph), 4.72 (AB d, 2H, J 12.0, OCH₂Ph), 4.71-4.64 (overlapping signals, 3H, OCH₂Ph, H-1), 3.95 (dd, 1H, J₂,₃ 2.0, J₃,₄ 11.0, H-3), 3.93-3.87 (overlapping signals, 3H, H-4,6a,6b), 3.76 (dd, 1H, J₁,₂ 2.5, H-2), 3.59 (dd, 1H, J₅, J 7.0, H-5), 3.31 (s, 3H, OMe), 1.12-1.04 (multiple signals, 21H, TIPS); ¹³C NMR (CDCl₃): ³δ 138.68, 138.61, 138.4 (each s, each aromatic C), 128.3, 128.2, 127.9, 127.67, 128.63, 127.5, 127.4 (each d, each aromatic CH), 98.5 (d, C-1), 80.3, 76.7, 74.9, 73.3 (each d), 75.1, 72.5, 72.1 (each t, each CH₂Ph), 63.2 (t, C-6), 54.4 (q, OMe), 18.0, 17.9 (each q, each TIPS CH₃), 12.3 (each d, each TIPS CH₂); LRMS: Found, 638.5 required, 638.9; [M + H₂O]+.

3.1.5 Methyl 2,3,4-tri-O-benzyl-6-O-triisopropylsilyl-α-D-galactopyranoside

Treatment of 1d (4.0 g, 20.0 mmol) as described for 1a gave 2d (6.4 g, 50%); [α]D 20.6º (c 0.07, CHCl₃); FTIR (KBr): 3030, 2865, 1496, 1454, 1350, 1194, 1054, 882, 793, 734, 696 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): ⁴δ 7.41-7.22 (multiple signals, 15H, each aromatic H), 4.82 (AB d, 2H, J 12.0, OCH₂Ph), 4.71 (AB d, 2H, J 11.5, OCH₂Ph), 4.77 (AB d, 2H, J 12.0, OCH₂Ph), 4.68 (d, 1H, J₁,₂ 3.5, H-1), 4.04 (dd, 1H, J₂,₃ 10.0, H-2), 3.95-3.92 (overlapping signals, 2H, H-3,5), 3.74-3.64 (overlapping signals, 3H, H-4,6), 3.36 (s, 3H, OMe), 1.12-0.86 (multiple signals, 21H, TIPS); ¹³C NMR (CDCl₃): ⁵δ 137.9, 137.7, 137.5 (each s, each aromatic C), 127.33, 127.28, 127.22, 127.15, 127.06, 126.62, 126.48, 126.45 (each d, each aromatic CH), 97.6 (d, C-1), 78.1, 75.4, 74.0, 70.1 (each d), 73.7, 72.5, 72.2 (each t, each CH₂Ph), 61.4 (t, C-6), 54.1 (q, OMe), 16.94, 16.93 (each q, each TIPS CH₃), 10.8 (each d, each TIPS CH₂); LRMS: Found, 638.5 required, 638.9; [M + H₂O]+.

3.1.6 Methyl 2,3,4-tri-O-benzyl-6-O-lauroyl-α-D-glucopyranoside (3a)
Compound 2a (3.0 g, 4.8 mmol) was dissolved in THF anhydrous (80 mL) and was cooled to 0 °C. Tetrabutyrammonium fluoride (1 g, 4 mmol) was added and the solution was allowed to warm to room temperature and stir for 1 h.²⁵ It was then concentrated in vacuo and approximately 1 mmol of the resulting 6-OH residue was dissolved in pyridine anhydrous (25 mL). 4-Dimethylaminopyridine and lauroyl chloride (0.29 mL, 1.22 mmol) were added and the solution was allowed to stir at room temperature for 24 h.²⁶ It was then concentrated under reduced pressure and the resulting benzylated ester derivative was purified by chromatography (petroleum ether-EtOAc) to give 3a (0.47 g, 72%); [α]D 7.5º (c 0.02, CHCl₃); FTIR (KBr): 2924, 2853, 1738, 1603, 1544, 1249, 1072 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.26 (ms, 15H, aromatic H), 4.92, (AB d, 2H, J 10.5, OCH₂Ph), 4.72, (AB d, 2H, J 10.5, OCH₂Ph), 4.64 (AB d, 2H, J 12.0, OCH₂Ph), 4.59 (d, 1H, J₁₂ 3.5, H-1), 4.27 (d, 2H, J₅,₆ 3.5, H-6a,6b), 4.01 (apt t, 1H, J₂,₃ 9.5, J₃,₄ 9.0, H-3), 3.82 (d apt t, 1H, J₄,₅ 10.0, H-5), 3.53 (dd, 1H, H-2), 3.48 (apt t, 1H, H-4) 3.37 (s, 3H, OCH₃), 2.35 (m, 2H, aliphatic OCOCH₂C₁₀H₂₁), 1.61 (m, 2H, aliphatic OCOCH₂CH₂C₉H₁₉), 1.28-1.24 (ms, 16H, aliphatic OCOC₂H₄C₅H₁₀CH₃), 0.87 (m, 3H, aliphatic OCOC₁₀H₂₀CH₃); ¹³C NMR (CDCl₃): δ 171.1 (s, C=O), 138.6, 138.1, 137.9 (each s, each aromatic C), 128.5, 128.48, 128.46, 128.1, 128.03, 127.98, 127.90, 127.7 (each d, each aromatic CH), 98.0 (d, C-1), 88.0, 79.9, 77.6, 68.6 (each d), 75.8, 75.1, 73.4 (each t, each CH₂Ph), 60.4 (t, C-6), 55.2 (q, OCH₃), 34.2, 31.9, 29.8, 29.6, 29.5, 29.3, 29.2, 24.9, 22.7, 21.1 (each t, each aliphatic CH₂), 14.2 (q, aliphatic CH₃); LRMS: Found, 669.39; required, 669.85; [M + Na]⁺; Anal. Calcd. for C₄₀H₅₄O₇: C, 74.27; H, 8.41. Found: C, 73.98; H, 8.30.

3.1.7 Methyl 2,3,4-tri-O-benzyl-6-O-lauroyl-β-D-glucopyranoside (3b)
Treatment of 2b (3.0 g, 4.8 mmol) as described for 2a gave 3b (2.2 g, 70%); [α]D 8.3°

(c 0.03, CHCl3); FTIR (KBr): 2924, 2853, 1739, 1497, 1454, 1356, 1151, 1070, 735 cm⁻¹; ¹H NMR (400 MHz, CDCl3): δ 7.36-7.24 (ms, 15H, aromatic H), 4.87, 4.84, 4.72 (each AB d, 6H, J 10.5, OCH₂Ph), 4.37 (d, 2H, J₅,₆ 11.5, H-6a,6b), 4.31 (d, 1H, J₅,₂ 8.0, H-1), 4.25 (m, 1H, H-5), 4.15 (t, 1H, J₂₃ 8.5, J₃,₄ 8.5, H-3), 3.56 (s, 3H, OCH₃), 3.54 (m, 1H, H-4), 3.43 (dd, 1H, H-2), 2.32 (m, 2H, aliphatic OCOCH₂C₁₀H₂₁), 1.62 (m, 2H, aliphatic OCOCH₂CH₂C₉H₁₉), 1.26-1.24 (ms, 16H, each aliphatic OCOCC₄H₄C₈H₁₆CH₃), 0.88 (t, 3H, J 6.0, J 7.0, aliphatic OCOC₁₁H₂₃CH₃); ¹³C NMR (CDCl₃): δ 173.6 (s, C=O), 138.43, 138.42, 137.8 (each s, each aromatic C), 128.8, 128.5, 128.4, 128.38, 128.34, 128.26, 128.11, 128.07, 127.97, 127.92, 127.8, 127.7, 127.69, 127.64, 127.5 (each d, each aromatic CH), 104.7 (d, C-1), 84.6, 82.3, 77.6, 72.9 (each d), 75.7, 75.1, 74.8 (each t, each OCH₂Ph), 62.9 (t, C-6), 57.1 (q, OCH₃), 34.2, 31.9, 29.6, 29.5, 29.3, 29.2, 29.1, 24.9, 24.7, 22.6 (each t, each aliphatic CH₂), 14.1 (q, aliphatic CH₃); LRMS: Found, 669.2 required, 669.9 [M + Na]⁺; Anal. Calcd. for C₄₀H₅₄O₇: C, 74.27; H, 8.41. Found: C, 73.91; H, 8.79.

3.1.8  Methyl 2,3,4-tri-O-benzyl-6-O-lauroyl-α-D-mannopyranoside (3c)

Treatment of 2c (6.2 g, 10.0 mmol) as described for 2a gave 3c (4.1 g, 64%); [α]D

23.3° (c 0.04, CHCl₃); FTIR (KBr): 3031, 2924, 2853, 1737, 1496, 1454, 1362, 1066, 1027, 970, 909, 735, 697 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.38-7.25 (multiple signals, 15H, each aromatic H), 4.77 (AB d, 2H, J 10.5, OCH₂Ph), 4.74 (d, 1H, J₅,₂ 2.0, H-1), 4.72 (AB d, 2H, J 12.5, OCH₂Ph), 4.61 (s, 2H, OCH₂Ph), 4.38 (dd, 1H, J₅,₆a 2.5, J₆₅₆b 12.0, H-6a), 4.33 (dd, 1H, J₅,₆b 5.0, H-6b), 3.94-3.88 (overlapping signals, 2H, H-3,4), 3.78 (dd, 1H, J₂₃ 2.5, H-2), 3.76 (m, 1H, H-5), 3.31 (s, 3H, OMe), 2.32 (t, 2H, J 7.5, J 7.5, aliphatic OCOCH₂C₁₀H₂₁), 1.61 (m, 2H, aliphatic
OCOCH$_2$CH$_2$C$_9$H$_{19}$, 1.31-1.54 (ms, 16H, aliphatic OCOC$_2$H$_4$C$_8$H$_{16}$CH$_3$), 0.91-0.86 (m, 3H, aliphatic OCOC$_{10}$H$_{20}$CH$_3$); $^{13}$C NMR (CDCl$_3$): $\delta$ 173.7, (s, C=O), 138.32, 138.21, 138.17 (each s, each aromatic C), 128.4., 128.38, 128.33, 128.05. 127.90, 127.76, 127.63, 127.23 (each d, each aromatic CH), 98.9 (d, C-1), 75.2, 74.6, 74.4, 69.9 (each d), 80.1, 72.6, 72.1 (each t, each CH$_2$Ph), 63.3 (t, C-6), 54.8 (q, OCH$_3$), 34.2, 33.9, 31.9, 29.61, 29.48, 29.44, 29.33, 29.27, 29.17, 29.07, 24.9, 24.7, 23.8, 22.7, 21.1 (each t, each aliphatic CH$_2$), 14.1 (q, aliphatic CH$_3$); LRMS: Found, 664.6 required, 664.9; [M + H$_2$O]$^+$; Anal. Calcd. for C$_{40}$H$_{54}$O$_7$: C, 74.27; H, 8.41. Found: C, 74.35; H, 8.25.

3.1.9 Methyl 2,3,4-tri-O-benzyl-6-O-lauroyl-\(\alpha\)-D-galactopyranoside (3d)

Treatment of 2d (5.7 g, 9.2 mmol) as described for 2a gave 3d (3.6 g, 60%); $[\alpha]_D$ 27.8$^\circ$ (c 0.09, CHCl$_3$); FTIR (KBr): 3030, 2924, 2853, 1738, 1496, 1454, 1350, 1099, 1049, 735, 696 cm$^{-1}$; $^1$H NMR (400 MHz, CDCl$_3$): $\delta$ 7.41-7.23 (multiple signals, 15H, each aromatic H), 4.83 (AB d, 2H, J 12.0, OC$_2$H$_2$Ph), 4.81 (AB d, 2H, J 11.5, OCH$_2$Ph), 4.77 (AB d, 2H, J 12.0, OCH$_2$Ph), 4.68 (d, 1H, J$_{1,2}$ 3.5, H-1), 4.16 (dd, 1H, J 7.5, J 11.5, H-4), 4.07-4.03 (overlapping signals, 2H, H-2,5), 3.94 (dd, 1H, J 3.0, J 10.0 H-6a), 3.86-3.84 (overlapping signals, 2H, H-3,6b), 3.35 (s, 3H, OMe), 2.23 (m, 2H, aliphatic OCOCH$_2$C$_{10}$H$_{21}$), 1.57 (m, 2H, aliphatic OCOCH$_2$CH$_2$C$_9$H$_{19}$), 1.31-1.18 (ms, 16H, aliphatic OCOC$_2$H$_4$C$_8$H$_{16}$CH$_3$), 0.88 (t, 3H, J 6.5, J 7.0, aliphatic OCOC$_{10}$H$_{20}$CH$_3$); $^{13}$C NMR (CDCl$_3$): $\delta$ 173.4 (s, C=O), 138.7, 138.4, 138.2 (each s, each aromatic C), 128.42, 128.36, 128.32, 128.11, 127.90, 127.75, 127.59, 127.51, 127.21 (each d, each aromatic CH), 98.7 (d, C-1), 78.9, 76.3, 74.9, 68.4 (each d), 74.6, 73.63, 73.54 (each t, each CH$_2$Ph), 63.3 (t, C-6), 55.3 (q, OCH$_3$), 34.1, 33.8, 31.9, 29.359, 29.45, 29.32, 29.26, 29.12, 24.9, 24.8, 22.7 (each t, each aliphatic CH$_2$), 14.1
(q, aliphatic CH$_3$); LRMS: Found, 664.6 required, 664.9; [M + H$_2$O]$^+$; Anal. Calcd. for C$_{40}$H$_{54}$O$_7$: C, 74.27; H, 8.41. Found: C, 74.67; H, 8.68.

3.1.10 Methyl 6-O-lauroyl-$\alpha$-D-glucopyranoside (4a)

Compound 3a (0.34 g, 0.2 mmol) was dissolved in EtOH (1 mL) and Pd-C (0.1 g) was added. The mixture was allowed to shake under hydrogen atmosphere of 2 psi until all protecting groups had been removed, as shown by TLC, to yield 4a. The suspension was filtered and concentrated in vacuo.$^{27}$ (0.17 g, 86%); [$\alpha$]$_D^20$ (c 0.02, CHCl$_3$); FTIR (KBr): 3734, 3445, 2955, 2924, 2850, 2359, 2341, 1728 cm$^{-1}$; $^1$H NMR (400 MHz, CDCl$_3$): $\delta$ 4.75 (d, 1H, J$_{1,2}$ 3.5, H-1), 4.33 (m, 2H, H-6), 3.75-3.73 (overlapping signals, 2H, H-3,5), 3.35 (apt t, 1H, J$_{3,4}$ 9.5, J$_{4,5}$ 9.5, H-4), 3.54 (dd, 1H, J$_{2,3}$ 9.5, H-2), 3.41 (s, 3H, OMe), 2.35 (t, 2H, J 7.5, aliphatic OCOCH$_2$C$_9$H$_21$), 1.63 (m, 2H, aliphatic OCOCH$_2$CH$_2$C$_9$H$_21$), 1.38-1.23 (ms, 16H, aliphatic OCOC$_2$H$_4$C$_8$H$_16$CH$_3$), 0.88 (t, 3H, J 7.0, aliphatic OCOC$_{10}$H$_{20}$CH$_3$); $^{13}$C NMR (CDCl$_3$): $\delta$ 174.2 (s, C=O), 99.4 (d, C-1), 74.1, 71.9, 70.4, 69.8 (each d), 63.5 (t, C-6), 55.2 (q, OCH$_3$), 34.2, 31.9, 29.66, 29.64, 29.5, 29.4, 29.3, 29.2, 24.9, 22.7 (each t, each aliphatic CH$_2$), 14.1 (q, aliphatic CH$_3$); LRMS: Found, 399.3 required, 399.5; [M + Na]$^+$; Anal. Calcd. for C$_{19}$H$_{36}$O$_7$: C, 60.61; H, 9.64. Found: C, 60.69; H, 9.83.

3.1.11 Methyl 6-O-lauroyl-$\beta$-D-glucopyranoside (4b)

Treatment of 3b (2.0 g, 3.0 mmol) as described for 3a gave 4b (0.86 g, 75%); [$\alpha$]$_D$ – 25.5° (c 0.05, CHCl$_3$); FTIR (KBr): 3421, 2921, 1744, 1703, 1016 cm$^{-1}$; $^1$H NMR (400 MHz, CDCl$_3$): $\delta$ 4.40 (d, 1H, J$_{1,2}$ 11.5, H-1), 4.28 (dd, 1H, J$_{2,3}$ 6.0, H-2), 4.21 (d, 2H, J$_{5,6}$ 7.5, H-6), 3.54 (s, 3H, OCH$_3$), 3.49 (m, 1H, H-3), 3.39-3.31 (overlapping signals, 2H, H-4,5), 2.34 (m, 2H, aliphatic OCOCH$_2$C$_{10}$H$_{21}$), 2.02 (s, 3H, OH), 1.62 (m, 2H, aliphatic OCOCH$_2$CH$_2$C$_9$H$_{19}$), 1.28-1.26 (ms, 16H, aliphatic OCOC$_2$H$_4$C$_8$H$_{16}$CH$_3$), 0.88 (t, 3H, J 6.5, aliphatic OCOC$_{10}$H$_{20}$CH$_3$); $^{13}$C NMR (CDCl$_3$): $\delta$ 174.2 (s, C=O),
1 103.6 (d, C-1), 76.5, 73.9, 73.4, 70.3 (each d), 63.6 (t, C-6), 57.0 (q, OCH₃), 34.2,
2 31.9, 29.61, 29.60, 29.5, 29.3, 29.2, 29.1, 24.9, 22.7 (each t, each aliphatic CH₂), 14.1
3 (q, aliphatic CH₃); LRMS: Found, 399.1 required, 399.5 [M + Na]⁺; Anal. Calcd. for
4 C₁₉H₃₆O₇: C, 60.61; H, 9.64. Found: C, 60.25; H, 9.91.
5 3.1.12 Methyl 6-O-lauroyl-α-D-mannopyranoside (4c)
6 Treatment of 3c (3.3 g, 5.0 mmol) as described for 3a gave 4c (1.4 g, 75%); [α]D
7 33.3° (c 0.01, CHCl₃); FTIR (KBr): 3421, 2923, 1736, 1466, 1197, 1057 cm⁻¹; ¹H
8 NMR (400 MHz, CDCl₃): δ 4.70 (s, 1H, H-1), 4.45 (br s, 1H, OH), 4.36 (d, 2H, J 4.0,
9 H-6), 3.96-3.92 (overlapping signals, 2H, OH, H-2), 3.78 (dd, 1H, J₂,₃ 2.5, J₃,₄ 9.0, H-
10 3), 3.71 (m, 1H, H-5), 3.62 (apt t, 1H, J₄,₅ 9.5, H-4) 3.36 (s, 3H, OMe), 2.35 (t, 2H, J
11 7.5, J 7.5, aliphatic OCOCH₂C₁₀H₂₁), 1.61 (m, 2H, aliphatic OCOCH₂CH₂C₉H₁₉),
12 1.29-1.25 (ms, 16H, aliphatic OCOH₂C₈H₁₆CH₃), 0.88 (t, 3H, J 6.5, J 7.0, aliphatic
13 OCO₁₀H₂₀(CH₃); ¹³C NMR (CDCl₃): δ 174.7 (s, C=O), 100.9 (d, C-1), 71.5, 70.5,
14 70.4, 67.7 (each d), 63.9 (t, C-6), 54.9 (q, OCH₃), 34.2, 31.9, 29.7, 29.6, 29.5, 29.4,
15 29.36, 29.34, 29.19, 24.9, 22.7 (each t, each aliphatic CH₂), 14.1 (q, aliphatic CH₃);
16 LRMS: Found, 377.3 required, 377.5; [M + H]⁺; Anal. Calcd. for C₁₉H₃₆O₇: C, 60.61;
17 H, 9.64. Found: C, 60.71; H, 9.53.
18 3.1.13 Methyl 6-O-lauroyl-α-D-galactopyranoside (4d)
19 Treatment of 3d (2.8 g, 4.4 mmol) as described for 3a gave 4d (1.4 g, 86%); [α]D
20 56.25° (c 0.01, CHCl₃); FTIR (KBr): 3250, 2918, 1741, 1467, 1194, 1025 cm⁻¹; ¹H
21 NMR (400 MHz, CDCl₃): δ 4.63 (apt t, 1H, J 6.5, J 5.0, OH-3), 4.57 (d, 1H, J 6.5,
22 OH-2), 4.55 (d, 1H, J₁,₂ 3.5, H-1), 4.13 (dd, 1H, J₅,₆a 8.0, J₆a,₆b 11.5, H-6a), 4.07 (dd,
23 1H, J₅,₆b 4.0, H-6b), 3.75 (dd, 1H, H-5), 3.68 (apt t, 1H, J₃,₄ 3.5, J₄,₅ 3.0, H-4), 3.58
24 (ddd, 1H, J₂,₃ 10.0, J₂,OH 16.5, H-2), 3.52 (m, 1H, H-3), 3.24 (s, 3H, OMe), 2.28 (t,
25 2H, J 7.5, aliphatic OCOCH₂C₁₀H₂₁), 1.63 (t, 2H, J 7.0, aliphatic
OCOCH₂CH₂C₉H₁₉, 1.28-1.23 (ms, 16H, aliphatic OCOCH₂C₆H₁₃CH₃), 0.85 (t, 3H, J 7.0, aliphatic OCOCH₁₀H₂₀CH₃); ¹³C NMR (CDCl₃): δ 178.2 (s, C=O), 104.8 (d, C-1), 74.9, 74.1, 73.7, 73.1 (each d), 68.8 (t, C-6), 59.8 (q, OCH₃), 38.9, 36.5, 34.2, 34.1, 33.9, 33.9, 33.7, 33.7, 33.5, 33.5, 30.4, 29.5, 27.3, (each t, each aliphatic CH₂), 18.9 (q, aliphatic CH₃); LRMS: Found, 399.3 required, 399.5; [M + Na]+; Anal. Calcd. for C₁₉H₃₆O₇: C, 60.61; H, 9.64. Found: C, 60.60; H, 9.88.

3.1.14 Methyl 2,3,4-tri-O-benzyl-6-O-octanoyl-α-D-glucopyranoside (5a)

Compound 2a (5.0 g, 8.5 mmol) was dissolved in THF anhydrous (150 mL) and was cooled to 0 °C. Tetrabutylammonium fluoride (2.2 g, 8.5 mmol) was added and the solution was warmed to room temperature and stirred for 1 h.²⁵ The mixture was then concentrated in vacuo and the resulting 6-OH residue was dissolved in pyridine anhydrous (100 mL). 4-Dimethylaminopyridine and octanoyl chloride (2.9 mL, 17 mmol) was added and the mixture was stirred at room temperature for 24 h.²⁶ The solution was then concentrated under reduced pressure and purified by chromatography (petroleum ether-EtOAc) to give 5a (3.9 g, 63%); [α]D 20.8° (c 0.07, CHCl₃); FTIR (KBr): 2927, 1738, 1497, 1454, 1360, 1163, 1093, 738, 697 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.37-7.26 (ms, 15H, aromatic H), 4.93, (AB d, 2H, J 10.5, OCH₂Ph), 4.74, (AB d, 2H, J 12.0, OCH₂Ph), 4.73 (AB d, 2H, J 10.5, OCH₂Ph), 4.60 (d, 1H, J₄,₅ 3.5, H-1), 4.28 (d, 2H, J₅,₆ 3.0, H-6), 4.01 (apt t, 1H, J₂,₃ 9.5, J₃,₄ 9.5, H-3), 3.81 (m, 1H, H-5), 3.54 (dd, 1H, H-2), 3.48 (dd, 1H, J₄,₅ 10.5, H-4), 3.37 (s, 3H, OCH₃), 2.31 (m, 2H, aliphatic OCOCH₂C₆H₁₃), 1.62 (m, 2H, aliphatic OCH₂CH₂C₆H₁₁), 1.30-1.05 (ms, 8H, aliphatic OC₆H₄C₄H₅CH₃), 0.87 (m, 3H, aliphatic OC₆H₁₂CH₃); ¹³C NMR (CDCl₃): δ 173.8 (s, C=O), 138.8, 138.3, 138.1 (each s, each aromatic C), 128.7, 128.6, 128.3, 128.29, 128.27, 128.3, 128.25, 128.20, 128.1 127.9 (each d, each aromatic CH), 98.3 (d, C-1), 82.2, 80.2, 77.8, 68.9 (each d),...
76.1, 75.3, 73.6 (each t, each OCH₂Ph), 63.1 (t, C-6), 55.4 (q, OCH₃), 34.4, 31.9,
29.2, 25.0, 22.8, 17.9 (each t, each aliphatic CH₂), 14.3 (q, aliphatic CH₃); LRMS:
Found, 613.4 required, 613.7; [M + Na]⁺; Anal. Calcd. for C₃₆H₄₆O₇: C, 73.19; H,
7.85. Found: C, 73.25; H, 7.61

3.1.15 Methyl 6-O-octanoyl-α-D-glucopyranoside (6a)

Treatment of 5a (3.6 g, 6.2 mmol) as described for 3a gave 6a (1.44 g, 73%); [α]D
27.9° (c 0.4, CHCl₃); FTIR (KBr): 3388, 2922, 1712, 1465, 1193, 1106, 724 cm⁻¹; ¹H
NMR (400 MHz, CDCl₃): δ 5.82 (s, 3H, each OH), 4.76 (d, 1H, J₁,₂ 3.5, H-1), 4.35 (d,
2H, J₅,₆ 4.0, H-6), 3.78-3.72 (overlapping signals, 2H, H-3,5), 3.54 (dd, 1H, H-2), 3.41 (s,
3H, OCH₃), 3.36 (dd, 1H, H-4), 2.35 (m, 2H, aliphatic COCH₂C₆H₁₃), 1.64 (t, 2H, J 7.0,
aliphatic COCH₂CH₂C₅H₁₁), 1.31-1.05 (ms, 8H, aliphatic COC₂H₄C₄H₆CH₃), 0.88 (t, 3H,
J 5.5, J 7.0, aliphatic COC₆H₁₂CH₃); ¹³C
NMR (CDCl₃): δ 179.5 (s, C=O), 99.4 (d, C-1), 74.1, 72.0, 69.7, 70.3 (each d), 63.4
(t, C-6), 55.3 (q, OCH₃), 34.1, 31.7, 31.6, 29.9, 28.9, 24.8 (each t, each aliphatic
CH₂), 14.1 (q, aliphatic CH₃); LRMS: Found, 343.1 required, 343.4; [M + Na]⁺; Anal.
Calcd. for C₁₅H₂₈O₇: C, 56.23; H, 8.81. Found: C, 56.47; H, 8.73.

3.1.16 Methyl 2,3,4-tri-O-paramethoxybenzyl-6-O-triisopropylsilyl-α-D-
glucopyranoside (7a)

A solution of 1a (5.0 g, 25.0 mmol) in DMF anhydrous (120 mL) was treated with
triisopropylsilyl chloride (15 mL, 75 mmol) and imidazole (5 g, 75 mmol) and
allowed to stir at room temperature for 24 h. The crude TIPS protected intermediate
was then concentrated in vacuo and the resulting residue dissolved in EtOAc. It was
then washed with 10% HCl, water, followed by sat. aq. NaHCO₃, and finally sat. aq.
NaCl, before being dried over anhydrous MgSO₄, and concentrated under reduced
pressure. The TIPS protected crude residue was then split in two and half was
dissolved in DMF anhydrous (30 mL) and THF anhydrous (20 mL). This solution was then added dropwise at 0 °C to a suspension of NaH (2.5 g, 62.5 mmol) in DMF anhydrous (10 mL) and THF anhydrous (7 mL), paramethoxybenzyl chloride (17 mL, 125 mmol) and tetrabutylammonium iodide (18.5 g, 50 mmol). This was stirred at approximately 10 °C for 30 min and then allowed to warm to room temperature and stir for 24 h. MeOH (50 mL) was added to quench the mixture which was stirred for 1 h. The solution was then concentrated under diminished pressure and dissolved in EtOAc. It was washed with water, dried over anhydrous MgSO₄, and concentrated in vacuo. The resulting residue was purified by chromatography (petroleum ether-EtOAc) to give 7a. (5.15 g, 59%); [α]D 11.6° (c 0.05, CHCl₃); FTIR (KBr): 3479, 102936, 2864, 1464, 1421, 1360, 1302, 883, 820, 760, 683, cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.34-6.73 (ms, 12H, aromatic H), 4.88 (AB d, 2H, J 10.5 OCH₂Ph), 4.78 (d, 1H, J₁,₂ 5.0, H-1), 4.75, 4.71 (each AB d, 2H, J 12.0 OCH₂Ph), 4.63 (m, 1H, H-2), 3.99 (apt t, 1H, J₃,₄ 9.0, J₄,₅ 9.0, H-4), 3.89 (m, 2H, H6), 3.77 (m, 9H, each PhOCH₃), 3.57-3.49 (overlapping signals, 2H, H-3,5), 3.39 (s, 3H, OCH₃); ¹³C NMR (CDCl₃): δ 159.6, 159.5, 159.4, 131.6, 131.4, 131.0 (each s, each aromatic C), 129.99, 129.93, 129.8, 114.13, 114.08, 113.6 (each d, each aromatic CH), 98.1 (d, C-1), 82.2, 80.2, 77.8, 72.1 (each d), 75.8, 74.9, 73.2 (each t, each OCH₂Ph), 63.1 (t, C-6), 55.47, 55.40, 55.36 (each q, each PhOCH₃), 55.0 (q, OCH₃), 18.27, 18.25 (each q, each TIPS CH), 12.3 (d, each TIPS CH); LRMS: Found, 733.3 required, 733.9 [M + Na]⁺.

3.1.17 Methyl 2,3,4-tri-O-paramethoxybenzyl-6-O-triisopropylsilyl-β-D-glucopyranoside (7b)

Treatment of 1b (4.5 g, 23.17 mmol) as described for 1a gave 7b (10.1 g, 61%); [α]D 4.8° (c 0.05, CHCl₃); FTIR (KBr): 2939, 1586, 1464, 883, 821, 760, 683, cm⁻¹; ¹H
NMR (400 MHz, CDCl$_3$): δ 7.30-6.84 (ms, 12H, aromatic H), 4.85, 4.80, 4.73 (each AB d, 2H, J 10.5, OCH$_2$Ph), 4.27 (d, 1H, J$_{1,2}$ 7.5, H-1), 3.95 (m, 1H, H-6a), 3.87 (dd, 1H, J$_{1,2}$ 10.5, OC$_3$H$_2$Ph), 4.27 (d, 1H, J$_{1,2}$ 7.5, H-1), 3.95 (m, 1H, H-6a), 3.87 (dd, 1H, J$_{4,5}$ 11.0, J$_{5,6}$ 4.5, H-5), 3.78 (m, 9H, PhOCH$_3$), 3.59 (m, 1H, H-3), 3.53 (s, 3H, OCH$_3$) 3.36 (apt t, 1H, J$_{2,3}$ 9.0, H-2), 3.29-3.24 (overlapping signals, 2H, H-4,6b), 1.10-1.04 (ms, 21H, TIPS);

$^{13}$C NMR (CDCl$_3$): δ 159.5, 159.4, 131.2, 131.1, 130.9, (each s, each aromatic C), 129.9, 129.8, 128.7, 114.1, 114.04, 114.01 (each d, each aromatic CH), 104.7 (d, C-1), 84.7, 82.6, 77.5, 76.2 (each d), 75.7, 74.9, 74.7 (each t, each OCH$_2$PH), 62.7 (t, C-6), 56.8 (q, OCH$_3$), 55.5 (each q, each PhOCH$_3$), 18.3, 18.2 (each q, each TIPS CH$_3$), 12.2 (d, each TIPS CH); LRMS: Found, 733.3; required, 733.9 [M + Na]+.

3.1.18 Methyl 2,3,4-tri-O-paramethoxybenzyl-6-O-dodecanyl-α-D-glucopyranoside (8a)

Compound 7a (4.0 g, 5.5 mmol) was dissolved in THF anhydrous (100 mL) and was cooled to 0 °C. Tetrabutylammonium fluoride (1.4 g, 5.5 mmol) was added and the solution was allowed to warm to room temperature and stir for 1 h. The mixture was then concentrated in vacuo, and the resulting 6-OH residue was dissolved in DMF anhydrous (100 mL). 1-chlorododecane (1.8 mL, 11 mmol) was added and the solution was cooled to 0 °C before NaH (0.11 g, 2.75 mmol) was added portion wise. The mixture was then allowed to warm to room temperature and was stirred for 24 h. MeOH (50 mL) was added to quench the solution which was stirred for 1 h. The crude PMB protected ether was then concentrated under diminished pressure and purified by chromatography (petroleum ether-EtOAc) to give 8a (1.89 g, 50%); [α]$_D$ – 8.6° (c 0.06, CHCl$_3$); FTIR (KBr): 2924, 2854, 1613, 1586, 1464, 1359, 1301, 1248, 1172, 1037, 820 cm$^{-1}$; $^1$H NMR (400 MHz, CDCl$_3$): δ 6.85-7.30 (ms, 12H, aromatic H), 4.92 (d, 1H, J$_{1,2}$ 10.5, H-1), 4.85 (AB d, 2H, J 10.5, OCH$_2$PhOCH$_3$), 4.74 (dd, 1H,
$J_{2,3}$ 9.5, H-2), 4.69, (AB d, 2H, $J$ 10.5, OCH$_2$PhOCH$_3$), 4.60 (AB d, 2H, $J$ 11.5

OCH$_2$PhOCH$_3$), 4.55 (apt t, 1H, $J_{3,4}$ 9.5, H-3), 3.95 (m, 1H, H-5), 3.80 (s, 9H, each

PhOCH$_3$), 3.53-3.37 (overlapping signals, 3H, H-4,6a,6b), 3.36 (s, 3H, OCH$_3$), 1.60

(m, 2H, aliphatic CH$_2$C$_{11}$H$_{23}$), 1.30-1.25 (ms, 20H, aliphatic CH$_2$C$_{10}$H$_{20}$CH$_3$), 0.89 (t,

3H, $J$ 7.0, aliphatic C$_{11}$H$_{20}$CH$_3$); $^{13}$C NMR (CDCl$_3$): $\delta$ 159.6, 159.5, 159.4, 131.3,

131.0, 130.6 (each s, each aromatic C), 130.0, 129.8, 129.6, 114.07, 114.05, 114.03

(each d, each aromatic CH), 98.5 (d, C-1), 82.1, 79.8, 77.7, 70.2 (each d), 75.7, 74.9,

73.3 (each t, each OCH$_2$Ph), 72.0 (t, aliphatic OCH$_2$C$_{11}$H$_{23}$), 69.5 (t, C-6), 55.5 (q,

PhOCH$_3$), 55.3 (s, OCH$_3$), 32.2, 29.94, 29.91, 29.89, 29.87, 29.84, 29.7, 29.5, 28.4

(each t, each aliphatic CH$_2$), 14.4 (q, aliphatic CH$_3$); LRMS: Found, 745.5; required,

745.9; [M + Na]$^+$; Anal. Calcd. for C$_{43}$H$_{62}$O$_9$: C, 71.44; H, 8.64. Found: C, 71.09; H,

8.73.

3.1.19 Methyl 2,3,4-tri-O-paramethoxybenzyl-6-O-dodecanyl-$\beta$-D-

glucopyranoside (8b)

Treatment of 7b (3.2 g, 4.5 mmol) as described for 7a gave 8b (0.55 g, 85%); $[\alpha]_D$ 2º

c 0.01, CHCl$_3$; FTIR (KBr): 2923, 2851, 1614, 1464.40, 1421, 1359, 1302, 1254,

1173, 1072, 813 cm$^{-1}$; $^1$H NMR (400 MHz, CDCl$_3$): $\delta$ 7.29-6.84 (ms, 12H, aromatic

H), 4.79, 4.75, 4.67 (each AB d, 2H, $J$ 10.5, OCH$_2$Ph), 4.26 (d, 1H, $J_{1,2}$ 7.5, H-1),

3.79-3.58 (overlapping signals, 2H, H-3,5), 3.79 (m, 9H, PhOCH$_3$), 3.68 (m, 2H, H-

6a,6b), 3.56 (s, 3H, OCH$_3$), 3.43-3.39 (overlapping signals, 2H, H-2,4), 1.63 (m, 2H,

aliphatic OCH$_2$C$_{11}$H$_{23}$), 1.29-1.24 (ms, 20H, aliphatic OCH$_2$C$_{10}$H$_{20}$CH$_3$), 0.88 (t, 3H,

$J$ 7.0, aliphatic OC$_{11}$H$_{22}$CH$_3$); $^{13}$C NMR (CDCl$_3$): $\delta$ 159.3, 159.2, 159.1, 130.9, 130.8,

130.5 (each s, each aromatic C), 129.8, 129.6, 129.5, 113.8, 113.7 (each d, each

aromatic CH), 104.8 (d, C-1), 84.4, 82.1, 77.7, 75.3 (each d), 74.9, 74.6, 74.4 (each t,

each OCH$_2$Ph), 71.9 (t, aliphatic CH$_2$), 69.7 (t, C-6), 57.1 (q, OCH$_3$), 55.3, 55.2 (each
3.1.20 Methyl 6-O-dodecanyl-α-D-glucopyranoside (9a)

Compound 8a (1.45 g, 2.0 mmol) was dissolved in a mixture of MeCN:H₂O (3:1) (21 mL) and ceric ammonium nitrate (8.85 g, 16.16 mmol) was added. The solution was allowed to stir at room temperature for 24 h. It was then concentrated in vacuo and purified by chromatography (petroleum ether-EtOAc) to give 9a (0.53 g, 73%); [α]D 78.8º (c 0.04, CHCl₃); FTIR (KBr): 3416, 2919, 2851, 1467, 1372, 1128, 1043, 1019 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 4.98 (br s, 1H, OH), 4.75 (d, 1H, J₁,₂ 3.5, H-1), 4.34 (br s, 1H, OH), 4.01 (br s, 1H, OH), 3.75 (apt t, 1H, J₂,₃ 9.5, J₃,₄ 9.5, H-3), 3.66 (m, 2H, H-6), 3.54-3.44 (overlapping signals, 3H, H-2,4,5), 3.37 (s, 3H, OCH₃), 1.58 (m, 2H, aliphatic CH₂C₁₁H₂₃), 1.28-1.25 (ms, 20H, each aliphatic CH₂C₁₀H₂₀CH₃), 0.88 (t, 3H, J₆.₅, J₇.₀, aliphatic CH₃); ¹³C NMR (CDCl₃): δ 99.7 (d, C-1), 74.5, 72.3, 72.2, 71.2 (each d) 70.6 (t, aliphatic CH₂), 69.5 (t, C-6), 55.4 (q, OCH₃), 32.1, 29.9, 29.88, 29.86, 29.83, 29.7, 29.6, 26.3, 22.9 (each t, each aliphatic CH₂), 14.3 (q, aliphatic CH₃); LRMS: Found, 385.2; required, 385.5; [M + Na]⁺; Anal. Calcd. for C₁₉H₃₈O₆: C, 62.95; H, 10.57. Found: C, 62.60; H, 10.67.

3.1.21 Methyl 6-O-dodecanyl-β-D-glucopyranoside (9b)

Treatment of 8b (0.44 g, 0.6 mmol) as described for 8a gave 9b (0.17 g, 76%); [α]D 1º (c 0.03, CHCl₃); FTIR (KBr): 3405, 2922, 2850, 1470, 1391, 1128, 1109, 1048 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 4.20 (d, 1H, J₁,₂ 7.5, H-1), 3.89 (s, 1H, OH), 3.74 (m, 2H, H-6a,6b), 3.66 (m, 1H, H-5), 3.54 (s, 3H, OCH₃), 3.52-3.44 (overlapping signals, 2H, H-3,4), 3.35 (apt t, 1H, J₂,₃ 8.0, H-2), 1.58 (m, 2H, aliphatic OCH₂C₁₁H₂₃), 1.28-1.11 (ms, 20H, aliphatic OCH₂C₁₀H₂₀CH₃), 0.88 (t, 3H, J 6.5, J
7.0, aliphatic OC$_{11}$H$_{22}$CH$_{3}$; $^{13}$C NMR (CDCl$_3$): $\delta$ 103.5 (d, C-1), 76.5, 74.4, 73.4, 72.1, (each d), 71.6 (t, aliphatic CH$_2$), 70.9 (t, C-6), 57.1 (q, OCH$_3$), 31.9, 29.7, 29.66, 29.65, 29.6, 29.58, 29.53, 29.4, 26.0, 22.7 (each t, each aliphatic CH$_2$), 14.1 (q, aliphatic CH$_3$); LRMS: Found, 385.2; required, 385.5; [M + Na]$^+$; Anal. Calcd. for C$_{19}$H$_{38}$O$_6$: C, 62.95; H, 10.57. Found: C, 62.83; H, 10.36.

3.1.22 Methyl 2,3-di-O-benzyl-4,6-di-O-benzylidene-α-D-glucopyranoside (10a)

A solution of 1a (1.0 g, 5.2 mmol), p-toluenesulfonic acid (10 mg) and benzaldehyde dimethylacetal (1.5 mL, 10.3 mmol) in acetonitrile anhydrous (25 mL) was stirred for 24 h at room temperature. Trimethylamine (0.5 mL) was added to neutralise the solution which was then stirred for 1 h. The product was filtered off as a white solid, washed with petroleum ether and dried. The benzylidene protected intermediate was then dissolved in DMF anhydrous (15 mL) and the solution was cooled to 0 °C. NaH (0.74 g, 18.4 mmol) was added slowly, followed by benzyl bromide (2.5 mL, 20 mmol). The mixture was then warmed to room temperature and stirred over night. MeOH (10 mL) was added to quench the solution which was stirred for a further 1 hr. The mixture was then concentrated under diminished pressure and purified by chromatography (petroleum ether-EtOAc) to give 10a. (2.0 g, 95%); [α]$_D$ 0.7º (c 0.05, CHCl$_3$); FTIR (KBr): 3063, 3031, 1109, 1088, 735, 692 cm$^{-1}$; $^1$H NMR (400 MHz, CDCl$_3$): $\delta$ 7.50-7.22 (ms, 15H, each aromatic H), 5.54 (s, 1H, CH$_2$Ph), 4.85 (AB d, 2H, J 4.0, OCH$_2$Ph), 4.82 (AB d, 2H, J 12.0, OCH$_2$Ph), 4.59 (d, 1H, $J_{1,2}$ 3.5, H-1), 4.26 (dd, 1H, $J_{5,6a}$ 10.0, $J_{6a,6b}$ 4.5, H-6a), 4.05 (apt t, 1H, $J_{2,3}$ 9.0, $J_{3,4}$ 9.0, H-3), 3.83 (m, 1H, H-5), 3.70 (apt t, 1H, $J_{5,6b}$ 10.5, H-6b), 3.62-3.54 (overlapping signals, 2H, H-2,4), 3.39 (s, 3H, OCH$_3$); $^{13}$C NMR (CDCl$_3$): $\delta$ 138.7, 138.1, 137.4 (each s, each aromatic C), 128.89, 128.43, 128.29, 128.20, 128.10, 127.90, 127.57, 126.0 (each d, each aromatic CH), 101.2 (d, C-1), 99.2 (d, CH$_2$Ph), 82.1, 79.2, 78.6, 62.3
(each d), 75.3, 73.8 (each t), 69.1 (t, C-6), 55.3 (q, OCH$_3$); LRMS: Found, 463.3 required, 463.5; [M + H]$^+$; Anal. Calcd. for C$_{28}$H$_{30}$O$_6$: C, 72.71; H, 6.54. Found: C, 72.31; H, 6.56.

3.1.23 Methyl 4,6-di-O-lauroyl-α-D-glucopyranoside (12a)

3.1.23.1 Methyl 2,3-di-O-benzyl-4,6-di-O-lauroyl-α-D-glucopyranoside (11a)

Compound 10a (1.7 g, 3.6 mmol) was dissolved in MeOH (50 mL) and a catalytic amount of TsOH was added. The solution was stirred at room temperature overnight, after which Et$_3$N (2 mL) was added to quench the reaction. The mixture was concentrated under diminished pressure and the crude diol residue was dissolved in pyridine anhydrous (70 mL). 4-Dimethylaminopyridine and lauroyl chloride (3.3 mL, 14.4 mmol) was added and the reaction was stirred at room temperature for 3 h. The solution was then concentrated under diminished pressure and purified by chromatography (petroleum ether-EtOAc) to give 11a. (1.0 g, 38%); FTIR (KBr): 2925, 2853, 1743, 1455, 1360, 1167, 1105, 1045, 734 cm$^{-1}$; $^1$H NMR (400 MHz, CDCl$_3$): δ 7.34-7.26 (multiple signal, 10H, each aromatic H), 5.01 (dd, 1H, $J_{3,4}$ 9.5, $J_{4,5}$ 10.0, H-4), 4.78 (AB d, 2H, $J$ 11.5, OCH$_2$Ph), 4.73 (AB d, 2H, $J$ 12.0, OCH$_2$Ph), 4.59 (d, 1H, $J_{1,2}$ 3.5, H-1), 4.15 (dd, 1H, $J_{5,6a}$ 5.5, $J_{6a,6b}$ 12.5, H-6a), 4.04 (dd, 1H, $J_{5,6b}$ 2.0, H-6b), 3.92 (apt t, 1H, $J_{2,3}$ 9.5, H-3), 3.87-3.82 (m, 1H, H-5), 3.56-3.5 (m, 1H, H-2), 2.36-2.27 (m, 4H, each aliphatic OCOCH$_2$C$_6$H$_5$), 1.67-1.56 (ms, 32H, each aliphatic OCOCH$_2$CH$_2$C$_9$H$_3$), 1.26-1.16 (ms, 32H, each aliphatic OCOCH$_2$CH$_2$C$_9$H$_3$), 0.88 (t, 6H, $J$ 6.5, $J$ 7.0, each aliphatic OCOC$_{10}$H$_2$OCH$_3$); $^{13}$C NMR (CDCl$_3$): δ 173.6, 172.4 (each s, each C=O), 138.4, 137.9 (each s, each aromatic C), 128.51, 128.32, 128.18, 128.05, 127.69, 127.57 (each d, each aromatic CH), 98.2 (d, C-1), 79.51, 79.18, 69.5, 67.7 (each d), 75.4, 73.6 (each t, each CH$_2$Ph), 62.2 (t, C-6), 55.4 (q, OCH$_3$), 34.15, 34.03, 33.99, 31.9, 29.62, 29.60, 29.49, 29.44,
3.1.23.2 Methyl 4,6-di-O-lauroyl-α-D-glucopyranoside (12a)

Compound 11a (0.84 g, 1.14 mmol) was dissolved in EtOH (2.5 mL) and Pd/C (0.3 g) was added. The mixture was allowed to shake under hydrogen atmosphere of 2 psi until all protecting groups had been removed as shown by TLC to yield 12a. The suspension was filtered and concentrated in vacuo. (0.47 g, 75%); [α]_D^4.33º (c 0.03, CHCl₃); FTIR (KBr): 3456, 2918, 2849, 1737, 1701, 1468, 1301, 1240, 1187, 81046 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 4.87 (dd, 1H, J₃,₄ 9.5, J₄,₅ 10, H-4), 4.82 (d, 1H, J₁,₂ 4.0, H-1), 4.23 (dd, 1H, J₅,₆b 2.0, J₆a,₆b 12.0, H-6b), 4.12 (dd, 1H, J₅,₆a 2.0, H-6a), 3.91 (ddd, 1H, H-3), 3.84 (apt t, 1H, J₂,₃ 9.5, H-3), 3.64 (m, 1H, H-2), 3.44 (s, 3H, OMe), 2.37-2.32 (m, 4H, each aliphatic OCOCH₂C₁₀H₂₁), 1.68-1.55 (m, 4H, each aliphatic OCOCH₂CH₂C₉H₁₉), 1.30-1.26 (multiple signals, 32 H, each aliphatic OCOC₂H₄C₈H₁₆CH₃), 0.88 (t, 6H, J 6.5, J 7.0, each aliphatic OCOC₁₀H₂₀CH₃); ¹³C NMR (CDCl₃): δ 173.63, 173.58 (each s, each C=O), 99.0 (d, C-1), 72.9, 72.7, 70.3, 67.7 (each d), 62.2 (t, C-6), 55.5 (q, OMe), 34.2, 34.1, 34.0, 31.9, 29.63, 29.61, 29.50, 29.47, 29.45, 29.36, 29.30, 29.27, 29.14, 29.08, 24.84, 24.82, 24.70, 22.70 (each t, each aliphatic CH₂), 14.1 (q, aliphatic CH₃); LRMS: Found, 559.5 required, 559.8; [M + H]⁺; Anal. Calcd. for C₃₁H₅₈O₈: C, 66.63; H, 10.46. Found: C, 66.66; H, 10.79.

3.1.24 General procedure for the preparation of pentaerythritol esters

Pentaerythritol 13 (1.0 g, 7.3 mmol), lauroyl chloride (4.8 mL, 21 mmol) and 4-dimethylaminopyridine were dissolved in pyridine anhydrous (50 mL) and stirred at 50 °C for 24 h. The solution was then concentrated in vacuo, and the following...
mono-lauroyl 14 and di-lauroyl 15 products were isolated by chromatography (petroleum ether-EtOAc) a tetra-lauroyl derivative was also isolated (0.39 g, 6%):

### 3.1.25 Mono lauroyl pentaerythritol (14)

(0.33 g, 14%); FTIR (KBr): 3462, 2914, 2848, 1737, 1712, 1476, 1187, 1038, 1005 cm\(^{-1}\); \(^1\)H NMR (400 MHz, CDCl\(_3\)): \(\delta\) 4.10 (s, 2H, \(CH_2OC=O\)), 3.80-3.61 (overlapping signals, 9H, 3 x \(CH_2OH\), 3 x OH), 2.34 (t, 2H, \(J\ 6.0, \ J\ 7.0,\ \) aliphatic \(OCOCCH_2\)), 1.61 (m, 2H, aliphatic \(OCOCCH_2CH_2C_9H_{19}\)), 1.26 (ms, 16H, aliphatic \(OCOCCH_2C_8H_{10}CH_3\)), 0.88 (m, 3H, aliphatic \(CH_3\)); \(^{13}\)C NMR (CDCl\(_3\)): \(\delta\) 175.0 (s, \(C=O\)), 62.7, 62.4 (each t, each \(CH_2OC\)), 45.3 (s, \(C(CH_2)_3\)), 34.2, 31.9, 29.59, 29.57, 29.44, 29.30, 29.23, 29.15, 24.9, 22.6 (each t, each aliphatic \(CH_2\)), 14.1 (q, aliphatic \(CH_3\)); LRMS: Found 341.2, required 341.45 \([M+Na]^+\); Anal. Calcd. for \(C_{17}H_{34}O_5\): C, 64.12; H, 10.76. Found: C, 64.08; H, 10.79.

### 3.1.26 Di lauroyl pentaerythritol (15)

(1.074 g, 29%); FTIR (KBr): 3351, 2915, 2850, 1739, 1701, 1471, 1163, 978, 719 cm\(^{-1}\); \(^1\)H NMR (400 MHz, CDCl\(_3\)): \(\delta\) 4.12 (s, 4H, each \(CH_2OC=O\)), 3.58 (s, 4H, each \(CH_2OH\)), 3.22 (br s, 2H, each OH) 2.34 (t, 4H, \(J\ 7.5, \ J\ 7.5,\ \) each aliphatic \(OCOCCH_2\)), 1.62 (t, 4H, \(J\ 6.5, \ J\ 6.5,\ \) each aliphatic \(OCOCCH_2CH_2C_9H_{19}\)), 1.26 (ms, 32H, each aliphatic \(OCOCCH_2C_8H_{10}CH_3\)), 0.88 (t, 6H, \(J\ 6.5, \ J\ 6.5,\ \) each aliphatic \(OCOC_{10}H_{20}CH_3\)); \(^{13}\)C NMR (CDCl\(_3\)): \(\delta\) 174.4 (s, each \(C=O\)), 62.4 (t, each \(CH_2OC\)), 44.7 (s, \(C(CH_2)_3\)), 34.2, 31.9, 29.56, 29.29, 29.21, 29.11, 24.9, 22.6 (each t, each aliphatic \(CH_2\)), 14.1 (q, each aliphatic \(CH_3\)); LRMS: Found 501.5, required 501.75 \([M+H]^+\); Anal. Calcd. for \(C_{29}H_{56}O_6\): C, 69.56; H, 11.27. Found: C, 69.64; H, 11.31.

### 3.1.27 Tetra lauroyl pentaerythritol
32 (0.39 g, 6%); FTIR (KBr): 2917, 2849, 1735, 1336, 1299, 1250, 1154, 1111, 1002 cm⁻¹

¹¹H NMR (400 MHz, CDCl₃): δ 4.11 (s, 8H, each CH₂OC=O), 2.30 (t, 8H, J 7.5, J 8.0, each aliphatic OCOCH₂CH₂C₉H₁₉), 1.60 (t, 8H, J 6.5, J 7.0, each aliphatic OCOCH₂CH₂C₉H₁₉), 1.41-1.26 (ms, 64H, each aliphatic OCOC₂H₄C₈H₁₆CH₃), 0.88 (t, 12H, J 6.5, J 7.0, each aliphatic OCOC₁₀H₂₀CH₃); ¹³C NMR (CDCl₃): δ 173.2 (s, each C=O), 62.1 (t, each CH₂O), 41.8 (s, C(CH₂)₄), 34.1, 31.9, 29.59, 29.45, 29.31, 29.23, 29.11, 24.8, 22.7 (each t, each aliphatic CH₂), 14.1 (each q, each aliphatic CH₃); LRMS: Found 888.7, required 888.36 [M+Na]⁺; Anal. Calcd. for C₅₃H₁₀₀O₈: C, 73.56; H, 11.65. Found: C, 73.60; H, 11.58.

3.2 Evaluation of anti-microbial activity

3.2.1 Preparation of bacterial cultures

Bacteria used in this study were *Staphylococcus aureus* ATCC 25923 and *Escherichia coli* ATCC 25922. Stock cultures were maintained in tryptic soy broth (TSB, Sharlau Chemie, Spain) supplemented with 20% glycerol at -70 °C. Cultures were routinely grown by subculturing 100 µL of stock culture into 9 mL TSB and incubating at 35 °C for 18 h. Cultures were then maintained on tryptic soy agar (TSA, Sharlau Chemie, Spain) plates at 4 °C. Working cultures were prepared by inoculating a loop of pure culture into TSB and incubating at 35 °C for 18 h. A bacterial suspension was prepared in saline solution (NaCl 0.85%, BioMérieux, France) equivalent to a McFarland standard of 0.5, using the Densimat photometer (BioMérieux, SA, France), to obtain a concentration of 1x10⁸ cfu/mL. This suspension was then serially diluted in TSB to obtain a working concentration of 1x10⁶ cfu/mL.

3.2.2 Anti-microbial activity assay

Stock solutions (100 mmol) of test compounds and standards were prepared in sterile hydroalcoholic diluent (ethanol-distilled water, 1:1) and stored at -20 °C. Stock
solutions were diluted in TSB to obtain initial working concentrations (10 or 20 mmol). Working test compounds and standards were serially diluted in sterile TSB to a final volume of 100 µL within the 96-well plate. 100 µL of freshly prepared inoculum of the organism under study was added to each appropriate well. The final concentration of each microorganism in each well was approximately 5x10^5 cfu/mL and the concentration range of chemical compounds was from 1:2 to 1:256. Each concentration was assayed in duplicate. The following controls were used in the microplate assay for each organism and test compound; blank: uninoculated media without test compound to account for changes in the media during the experiment; negative control: uninoculated media containing only the test compound; positive control 1: inoculated media without compound; positive control 2: inoculated media without compound but including the corresponding sugar to evaluate any effect of the sugar alone; and positive control 3: inoculated media without compound but with the equivalent concentration of ethanol used to dissolve the test compound, thereby assessing any activity of the alcohol. The 96-well plates were incubated at 35 °C for 18 hours in a microtiterplate reader (PowerWave microplate Spectrophotometer, BioTek) and effects were monitored by measuring the optical density (OD) at 600 nm for each well every 20 minutes with 20 seconds agitation before each OD measurement. Each experiment was replicated three times. The MIC was defined as the lowest concentration of compound that showed no increase in OD values for all the replicates compared to the negative control after 18 hours. Subtraction of the absorbance of the negative control eliminated interferences due to variation in the media.

Acknowledgements

We would like to thank Geraldine Fitzpatrick, Dr Ken Glass and Dr Yannick Ortin,
1 NMR Center UCD, and Dr Dilip Rye, Mass Spectroscopy Centre UCD for spectra.

2 Funding for this project was provided by TSR Strand I Grant.